



Challenges for Intelligent Image Processing in Electron Microscopy

Christoph Best

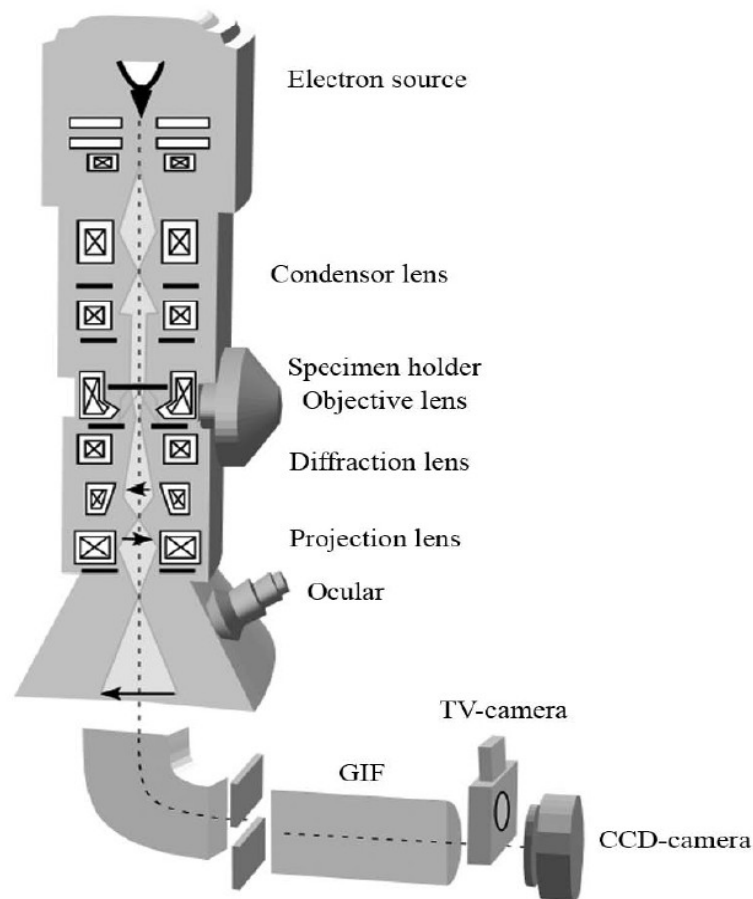
Max Planck Institut für Biochemie

Martinsried, Germany

Outline

- ▷ Introduction
- ▷ Tomography
 - ▷ Alignment belief networks
 - ▷ Reconstruction level sets; Monte Carlo algorithms
 - ▷ Denoising/segmentation anisotropic diffusion; scaling index
- ▷ Single-particle analysis EM algorithm
 - ▷ Automated particle picking supervised learning using SVMs
 - ▷ Model-free initial models unsupervised learning
probabilistic modelling

Electron microscopy



(Macro-)Molecular resolution $O(5 \text{ nm})$

Near-native state:

Cryo-electron microscopy:

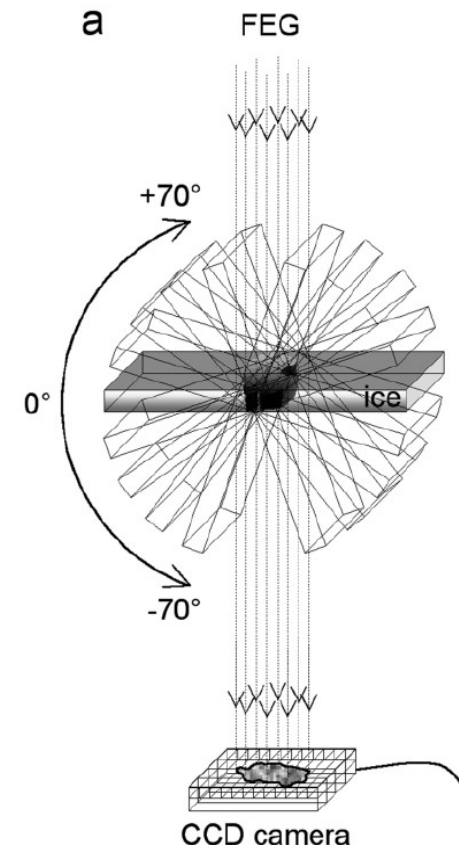
Specimen embedded in **vitreous ice**

Problems:

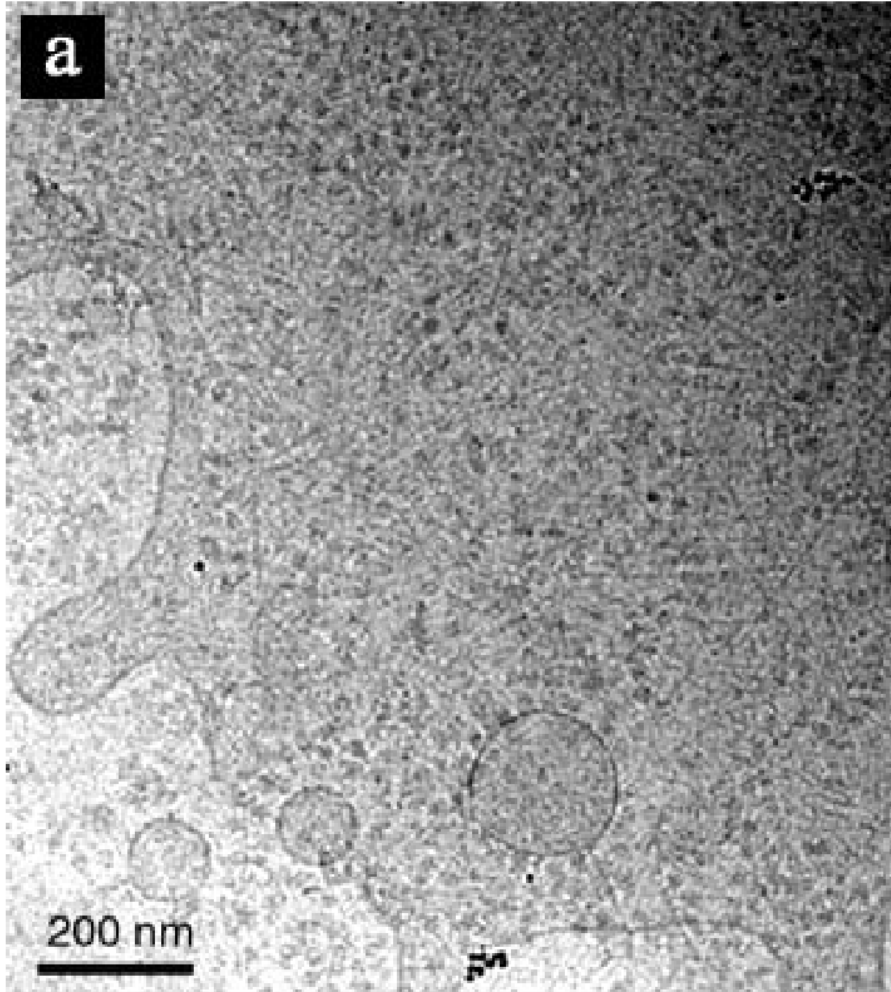
- Beam damage, low dose, high noise
→ **denoising**
- No fluorescent markers
→ **nano-gold markers**
- Low penetration (100nm)
→ **cryosectioning**

Electron tomography

- ▷ 3D reconstruction by taking a series of images from different angles
- ▷ Difficulty:
 - ▷ Nanometer accuracy (basically solved, at least for single axis)
- ▷ Problems:
 - ▷ Limited tilt range
→ **missing wedge**
 - ▷ Imperfections of the tilt
→ **alignment**
 - ▷ Reconstruction algorithms under high noise

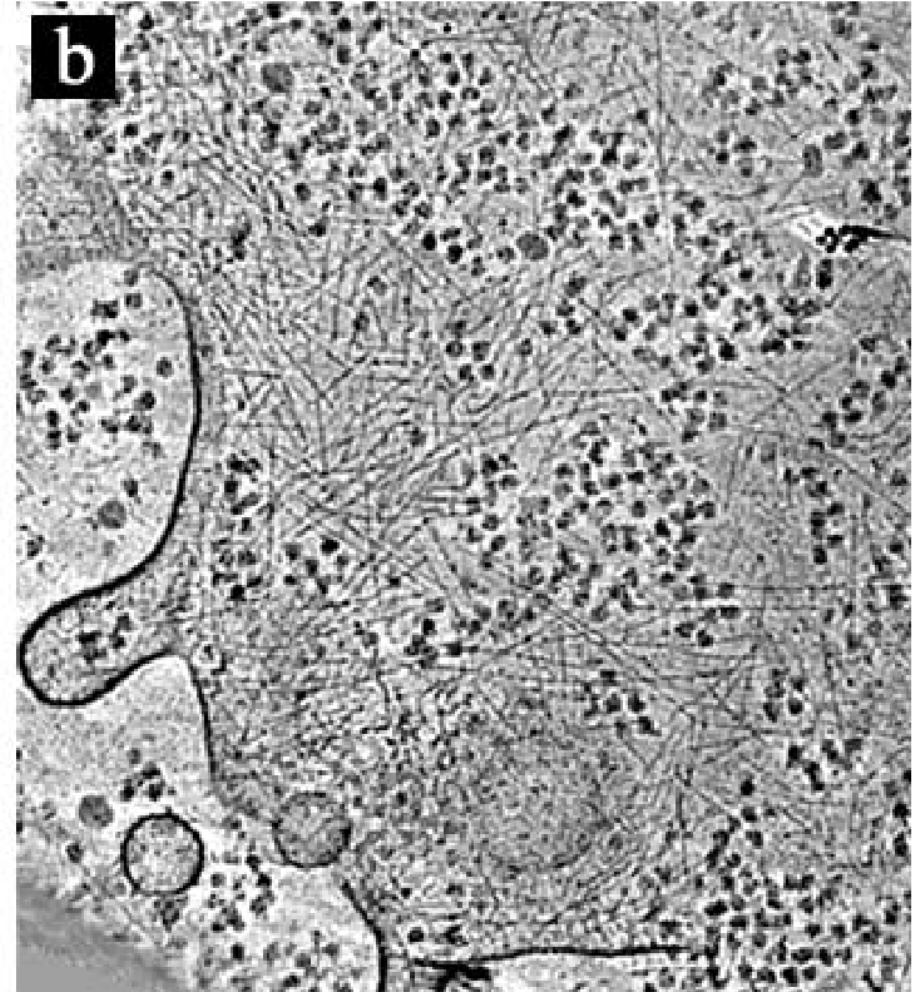


Tomography of eukaryotic cells



PROJECTION

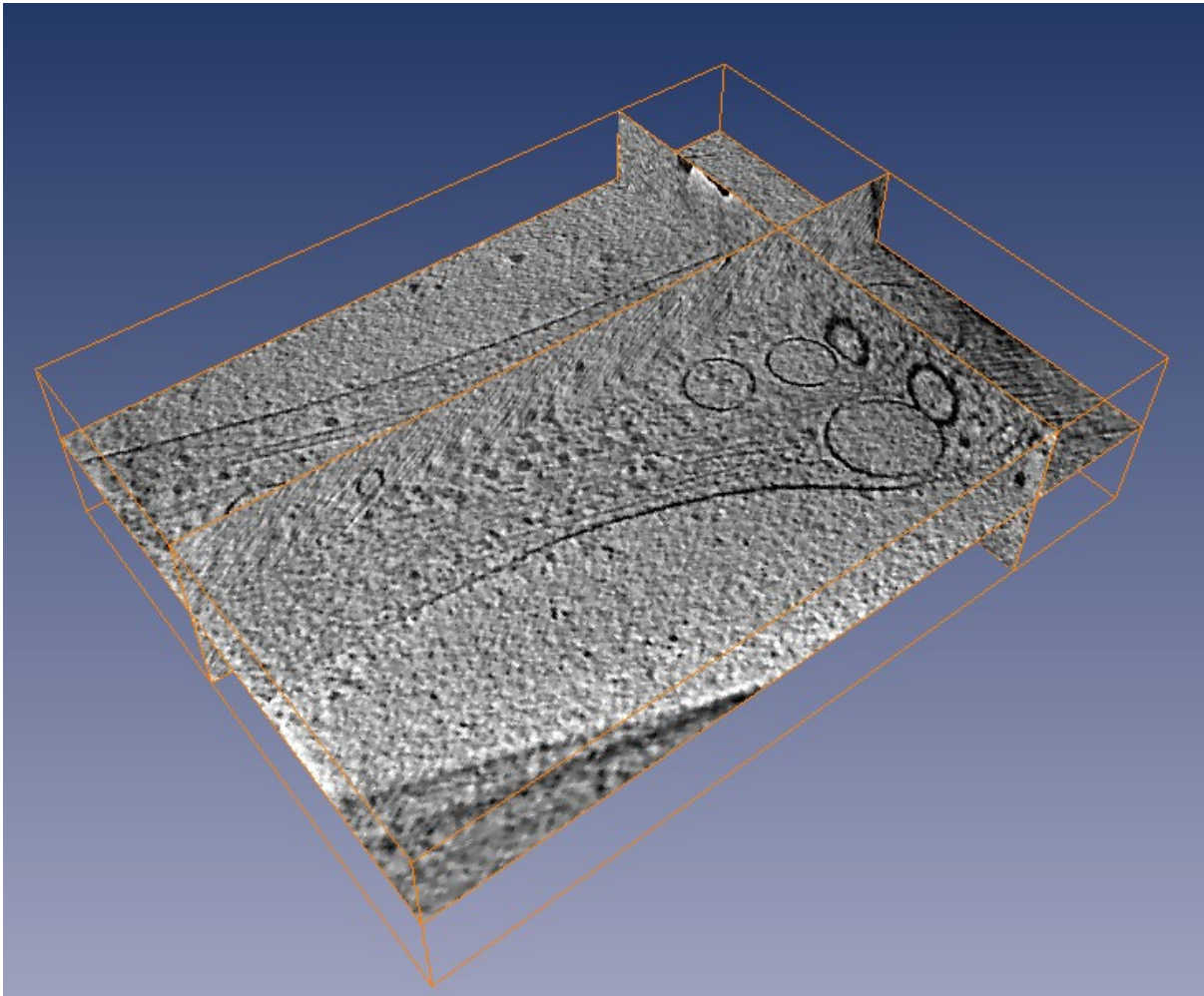
Dictyostelium discoideum



SLICE

O. Medalia et al, *Science*, 2002

3D imaging on a molecular scale



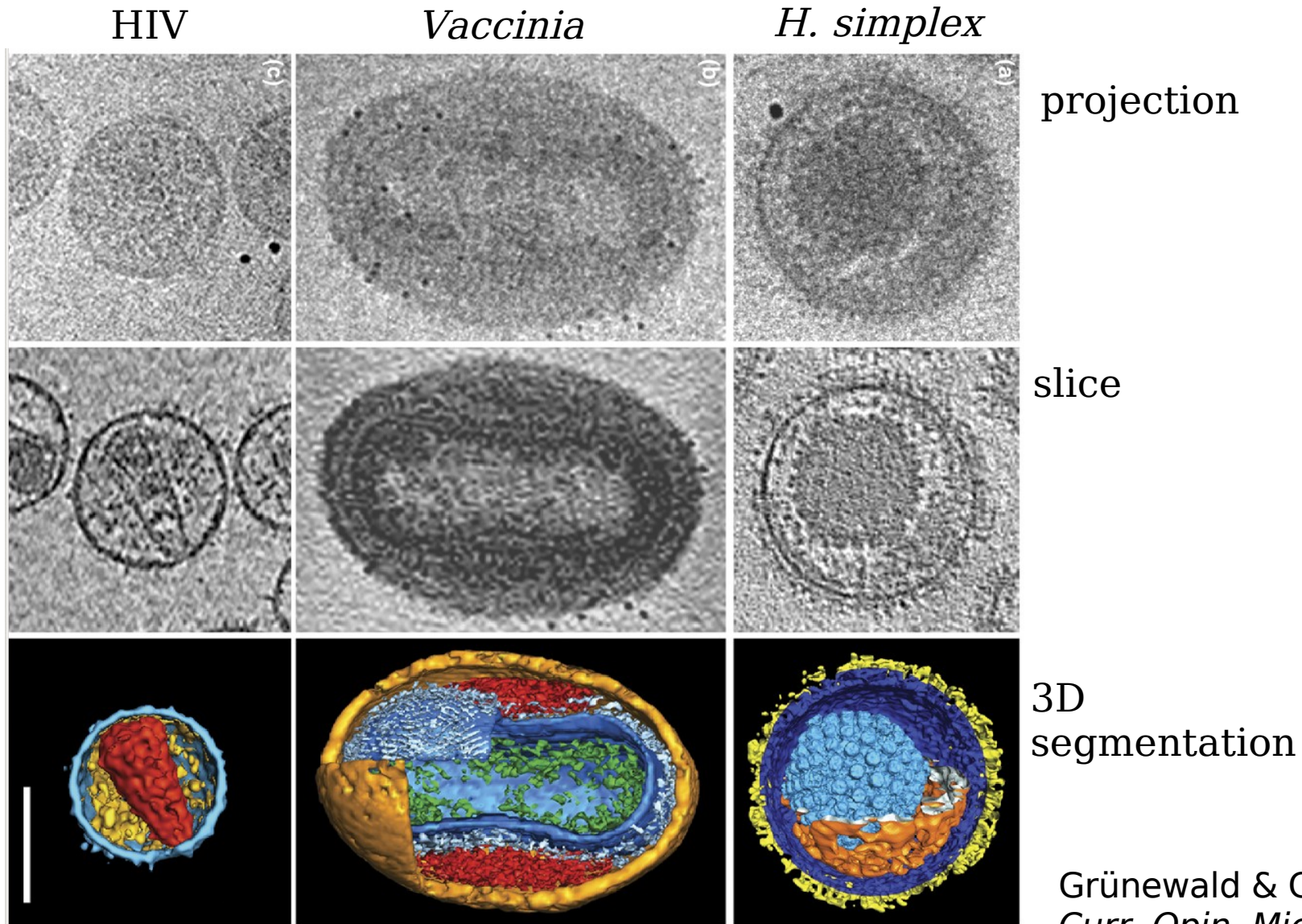
Mouse adeno-
carcinoma cell
grown on a
TEM grid

assembled from
66 images over
a 132° range

thickness $\approx 520\text{nm}$

courtesy of Andrew Leis, Martinsried

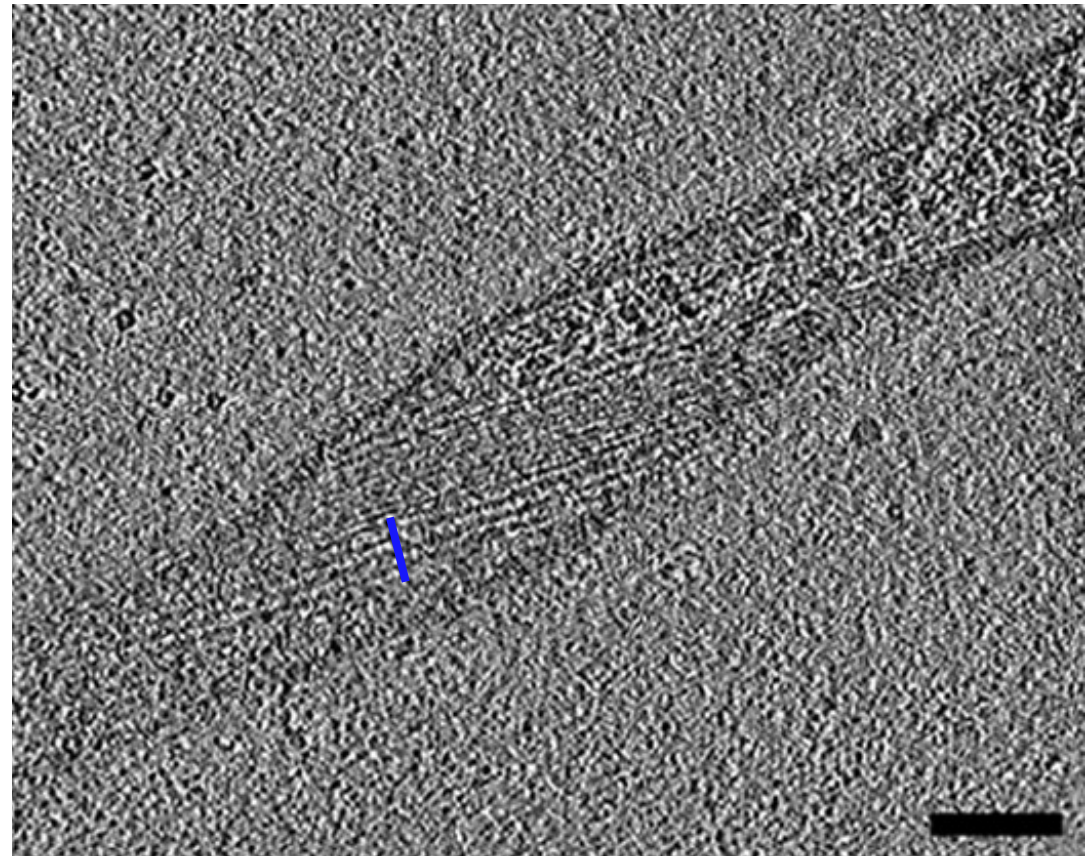
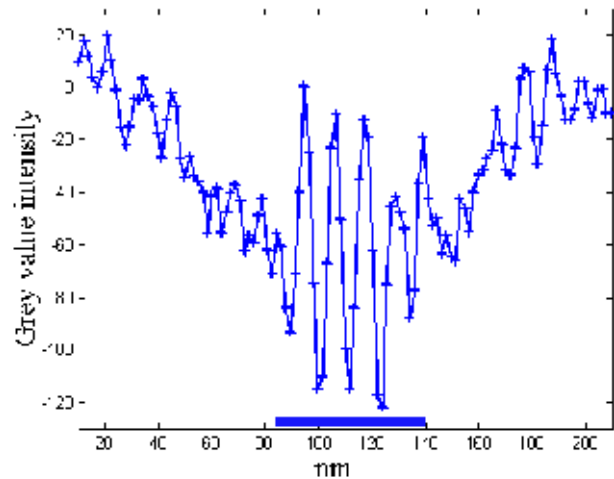
Viruses



Grünewald & Czirklaff,
Curr. Opin. Microbiol., 2006

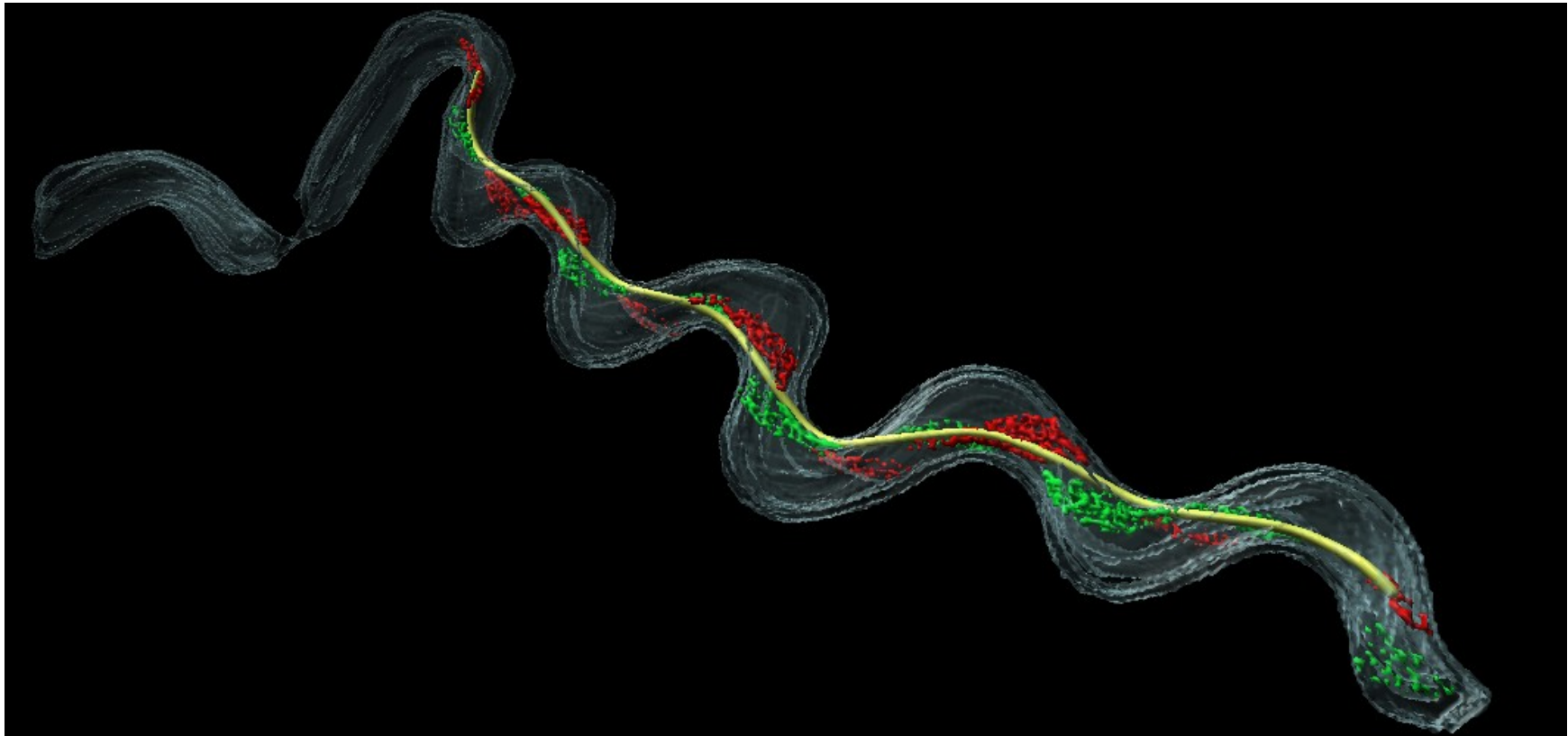
Cytoskeleton

Cytoskeleton of *Spiroplasma melliferum*



J. Kürner *et al.*, *Science*, 2005

The cytoskeleton of *S. melliferum*

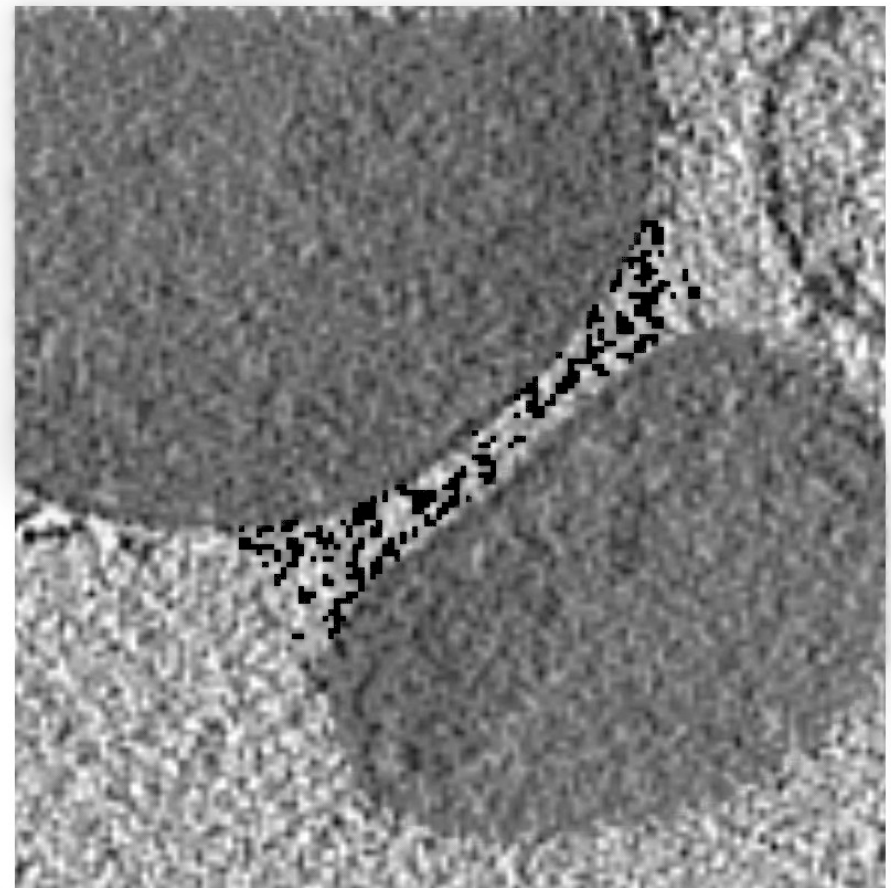
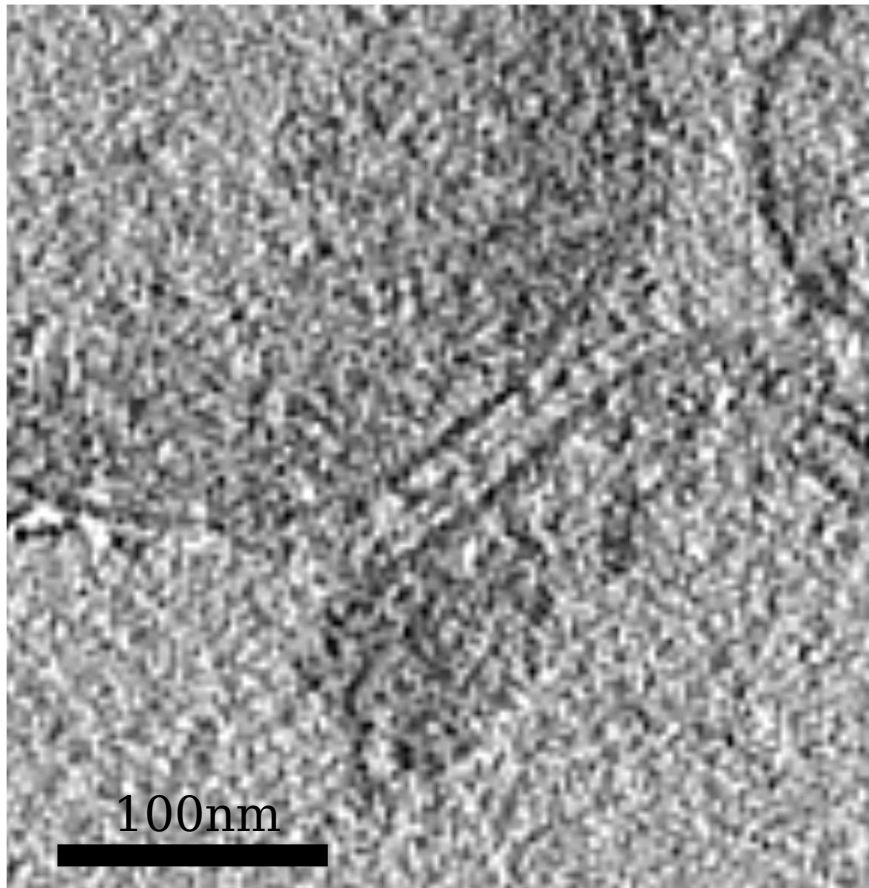


yellow: geodetic line

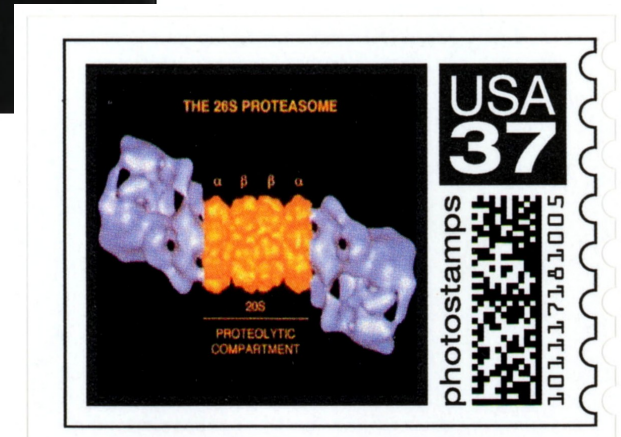
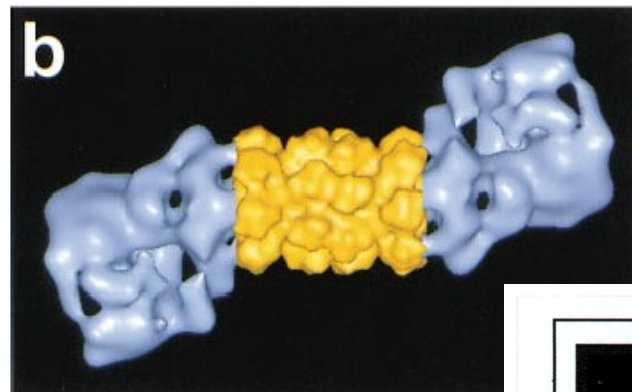
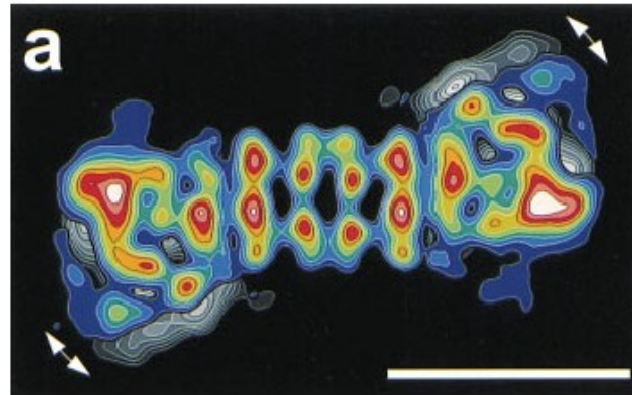
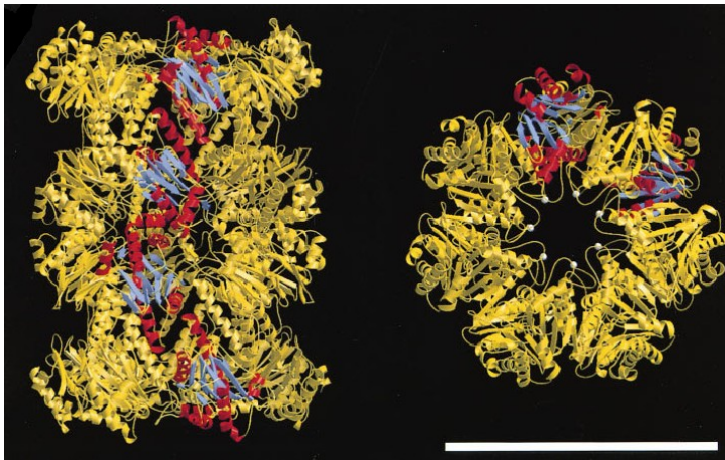
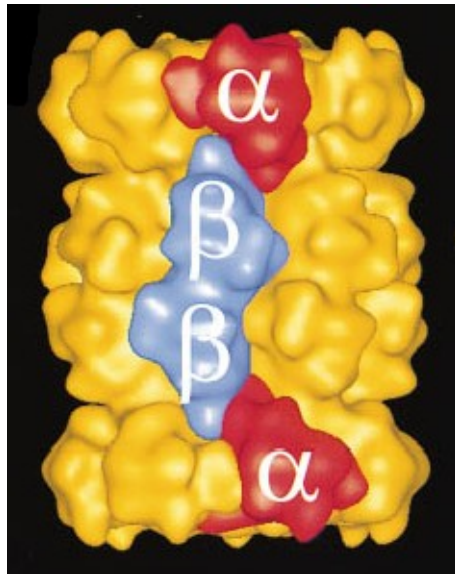
J. Kürner *et al.*, *Science*, 2005

Neurobiology applications

Molecules released in the synaptic cleft of a mammalian synapse.
(courtesy of V. Lucic, Martinsried).



Structural Biology: The Proteasome

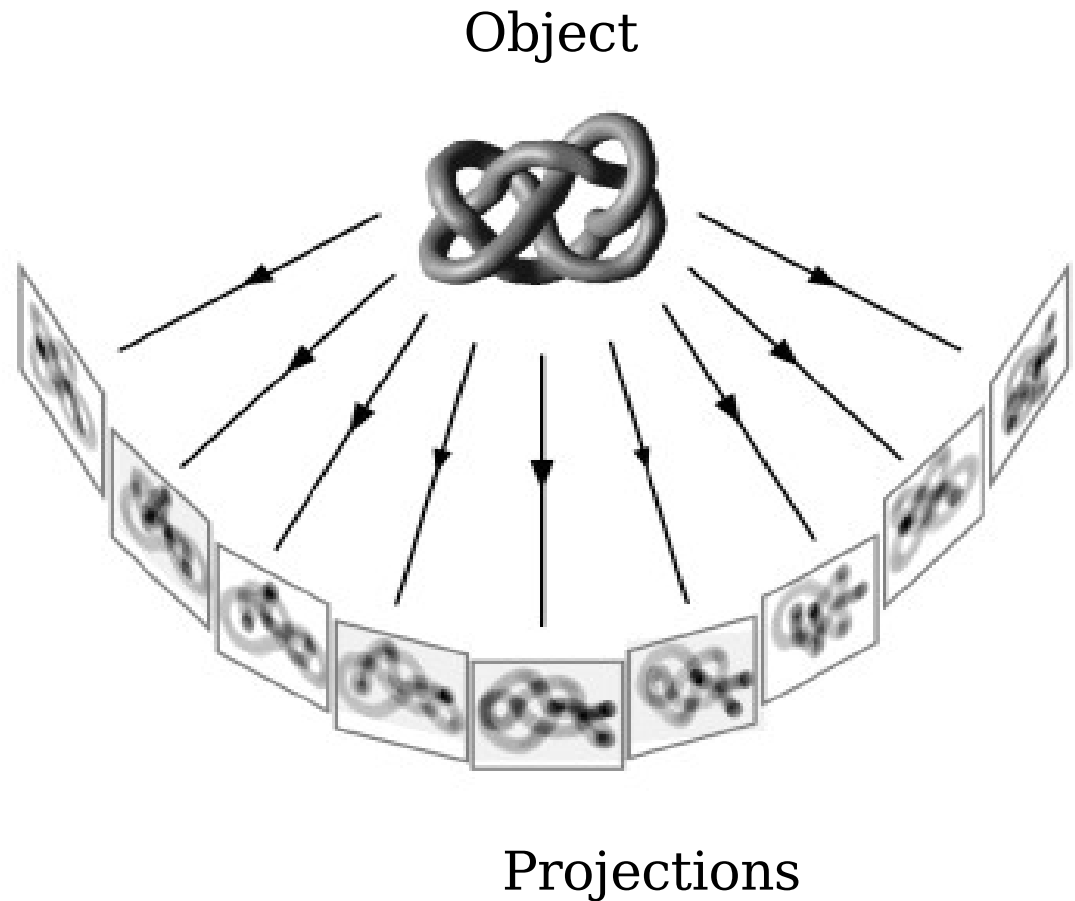


Tomography

Tomographic
reconstruction
problem:

**Find an object
consistent
with all projections.**

Data set size: \approx 500 MB
Acquisition time: 30 min



Alignment

- ▷ Problem:
Different images must be **shifted/rotated** to correspond to a common reference frame
- ▷ Strategies:
 - ▷ **Markers**: Add nanogold particles to object, use markers as reference points
[manual selection]
 - ▷ **Features**: Select features that can be identified in several images
 - ▷ **Correlation**: for images with overall similarity (based on a Gaussian noise model)

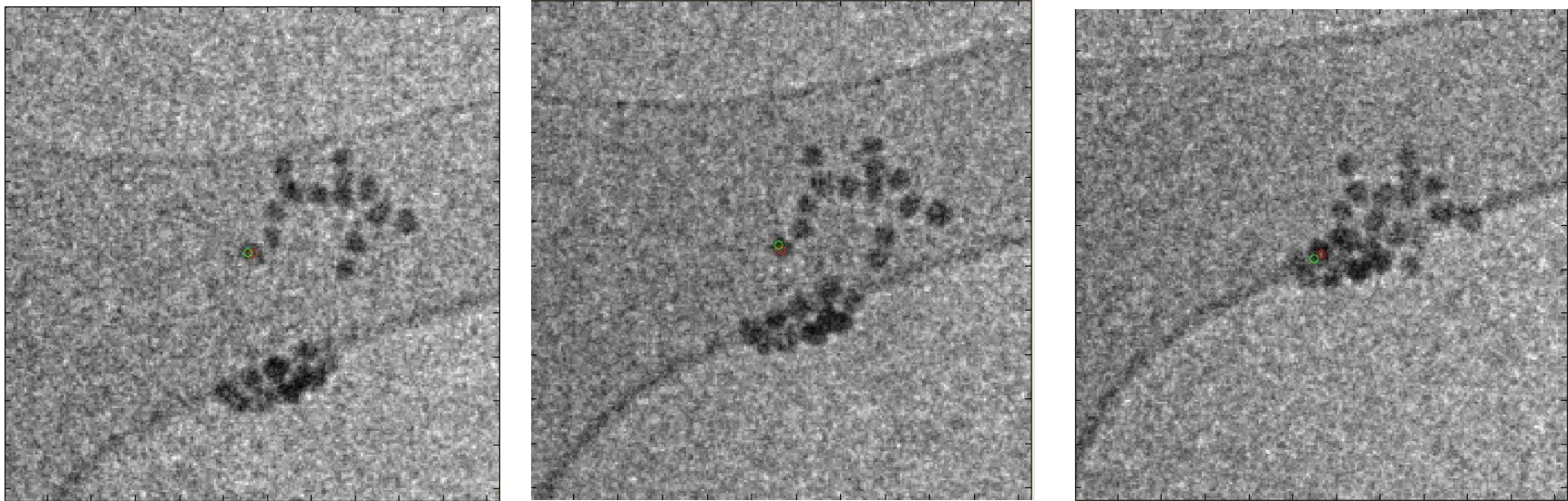
Algorithmic alignment problem

- ▷ Given:
projected positions $x^{(n)}(\phi^{(n)})$ taken at angles $\phi^{(n)}$
- ▷ Determine:
Which marker corresponds to which spots in the different projection images?
- ▷ **Difficulty:** Combinatorial problem
but there are good heuristics
- ▷ Applications:
 - ▷ Automatic alignment
 - ▷ Multiscopic immunomarker registration

Automated tracking

Fernando Amat, Farshid Moussavi, Mark Horowitz:

“Automatic tracking of fiducial markers across very low SNR images”, 4ICET, San Diego, 2006



Using a Markov Random Field and Loopy Belief Propagation

Tomographic reconstruction

- ▷ Classical result: Radon 1917

n-dimensional functions can be reconstructed from their (n-1)-dimensional integrals

- ▷ Special case: tomography:

integrals (=sums over voxels) along straight rays

- ▷ But how?

- ▷ Simplest algorithm: weighted back projection
- ▷ Iterative algorithms: ART, SIRT

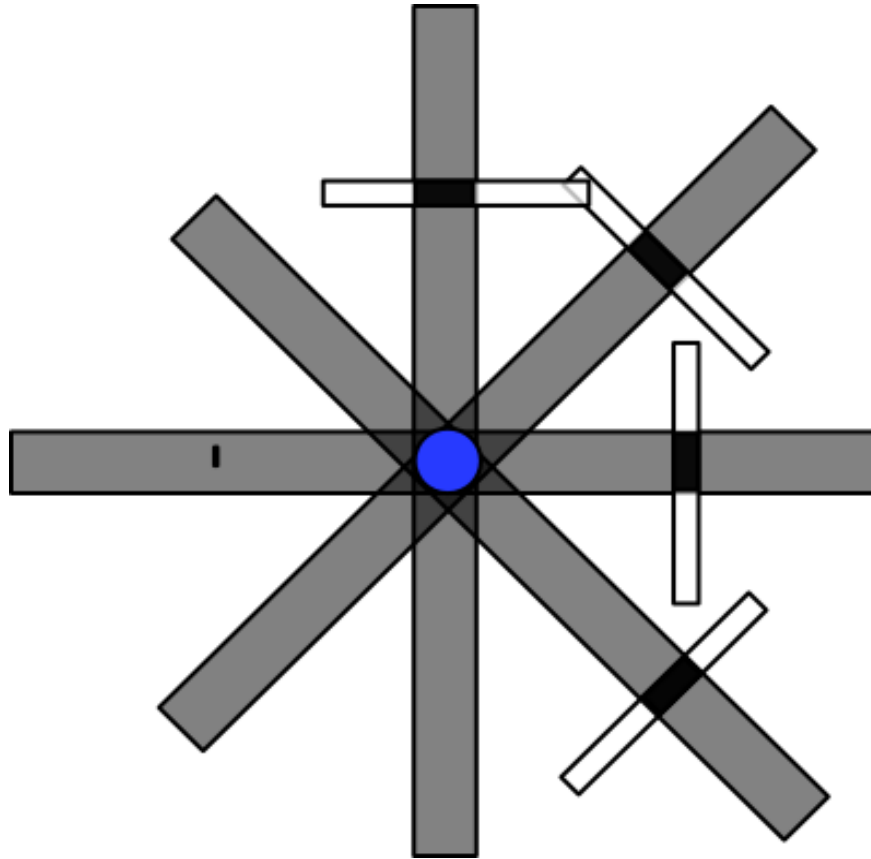
Reconstruction algorithms

▷ **Reconstruction problem:**

Given a set of projections and their angles, determine the/a 3D density that generates these projections

- ▷ Underdetermined problem (less image pixels than voxels in volume) → needs regularization
- ▷ Volumes are large: typically 512x512x256 or larger (several 100 MB to GB)
- ▷ Classical algorithms are simple: WBP, ART, SIRT

Weighted back projection

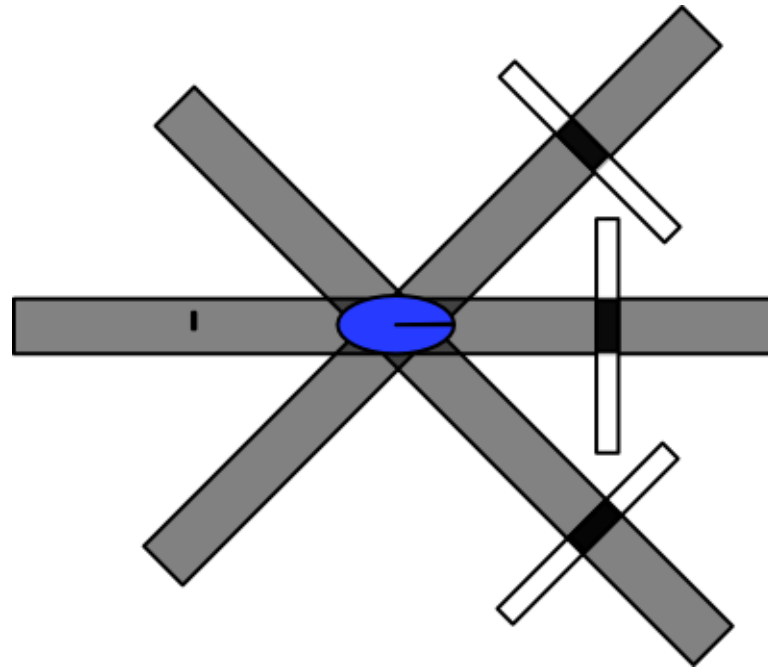


Computationally
very simple

First-order
approximation
to true solution

Good enough
when many
equally spaced
projections are
taken

Weighted back projection



Missing wedge → distortion

Iterative methods

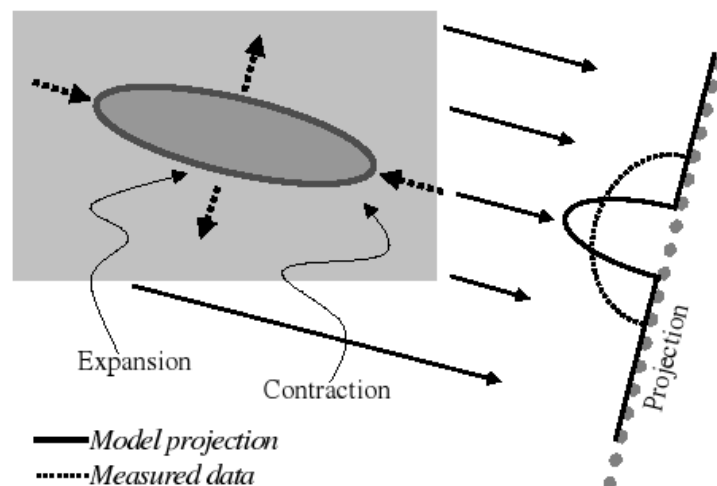
- ▷ Iteratively compare observed and calculated projections
- ▷ Update 3D density accordingly
- ▷ Classical implementations:
 - ▷ ART (**algebraic reconstruction technique**)
projections views processes sequentially
 - ▷ SIRT (**simultaneous iterative reconstruction**)
all projections processed in parallel
- ▷ Maximum-entropy methods (Skoglund, *JSB*, 1996)

Room for Improvement

- ▷ Iterative algorithms:
 - ▷ Conjugate-gradient
- ▷ Probabilistic methods:
 - ▷ Monte Carlo reconstruction
- ▷ Long-term goal: **Incorporate previous knowledge**
 - ▷ Discrete tomography
 - ▷ Combine denoising & reconstruction
 - ▷ Algorithms that require fewer projections

Level set reconstruction

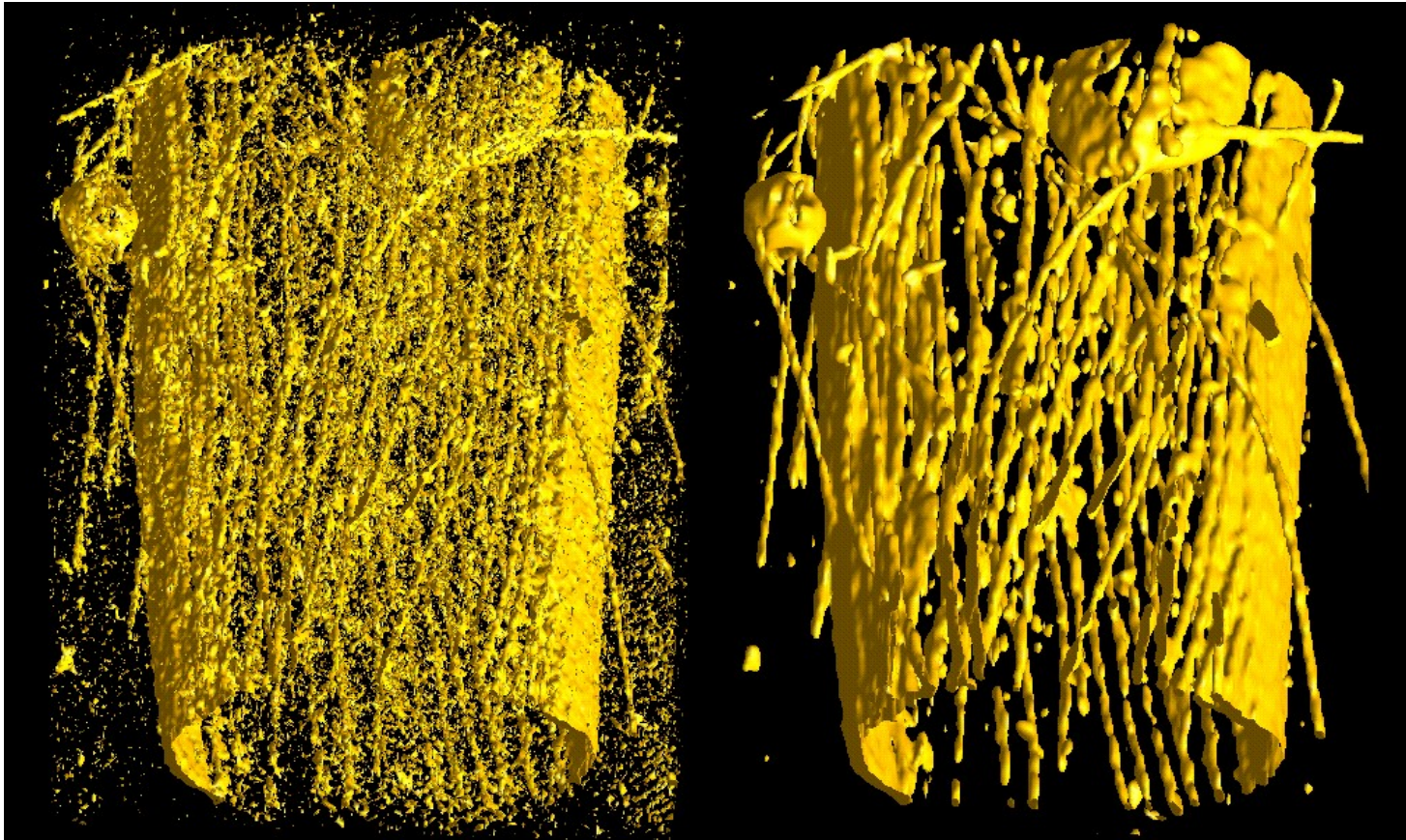
- ▷ Goal: reconstruct with a small number of views
- ▷ Idea:
 - ▷ Represent shape by a level set function
 - ▷ Compare projected outlines to views
 - ▷ Accumulate force from each view
 - ▷ Update level set
 - ▷ Combines segmentation with reconstruction



Anisotropic diffusion

- ▷ Why denoising?
 - ▷ Not for solving structures!
 - ▷ Important for **segmentation**
 - ▷ Useful for **visualization**
- ▷ Physical approach:
 - ▷ Diffusion is smoothing
 - ▷ Modify diffusion to respect edges
 - anisotropy and nonlinearity
- ▷ Diffusion constant is determined by local gradient

Anisotropic diffusion



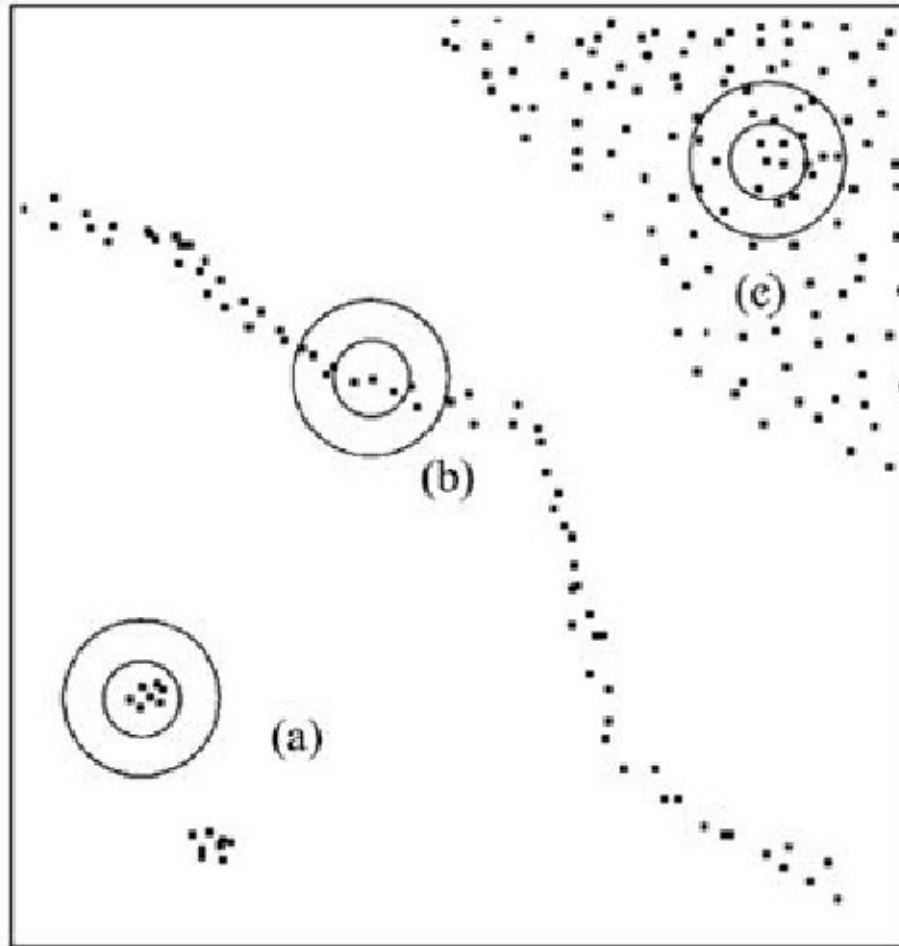
original

denoised

Electron tomographic reconstruction of a DMPC vesicle with actin filaments

R. Grimm, M. Bärmann, W. Häckl, D. Typke, E. Sackmann, W. Baumeister: *Biophys. J.* 72 (1997) 482-489

Scaling-index method



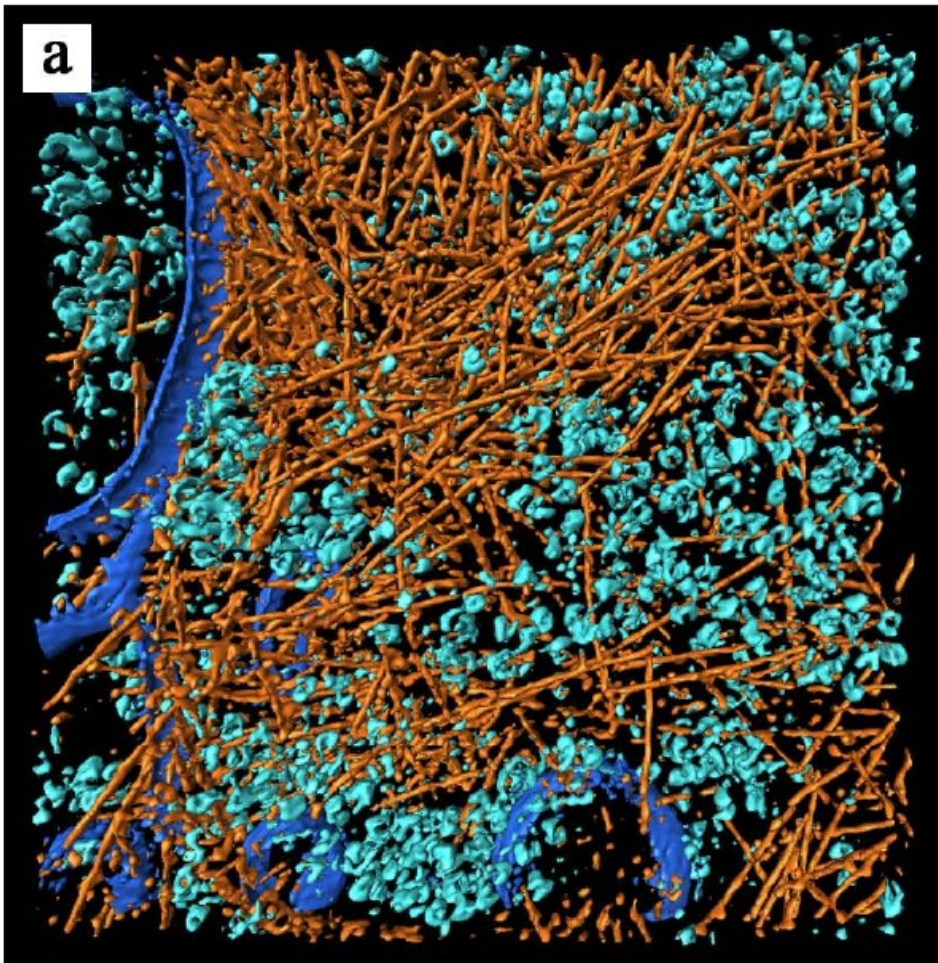
of points in a sphere
that belong to the object

characterizes the
dimensionality of the
object

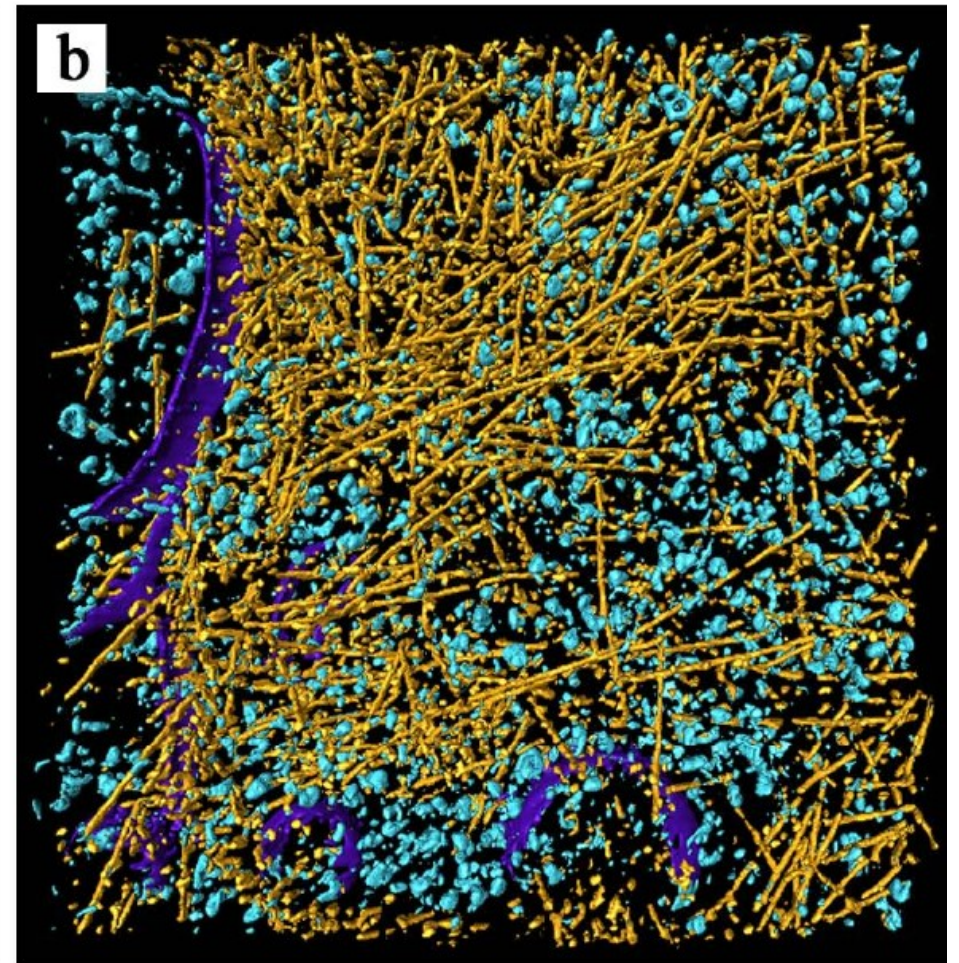
$$N(p_i, r) \propto r^\alpha$$

Scaling-index segmentation

A. Linaroudis, Ph.D. Thesis, 2006

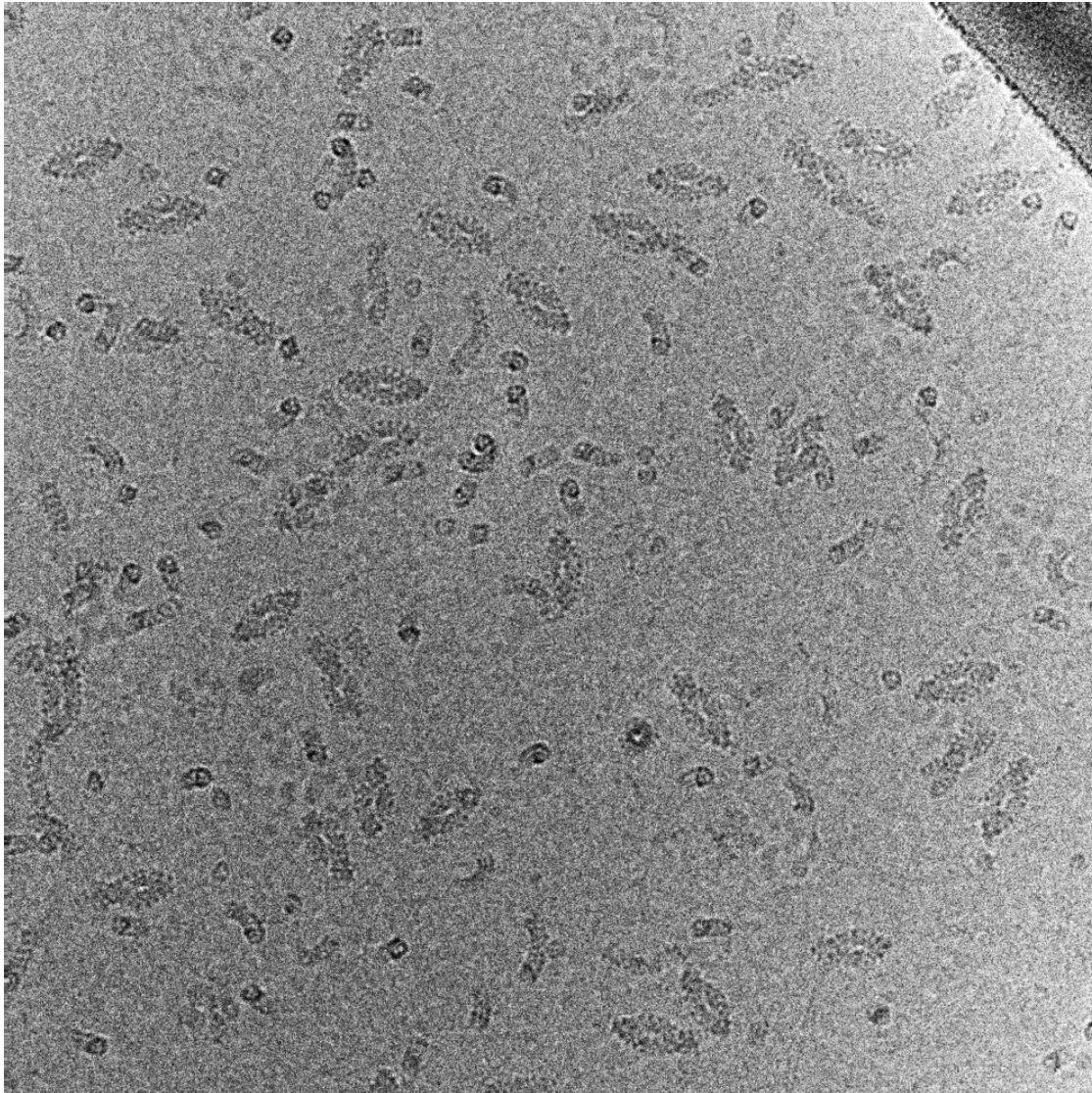


Manual



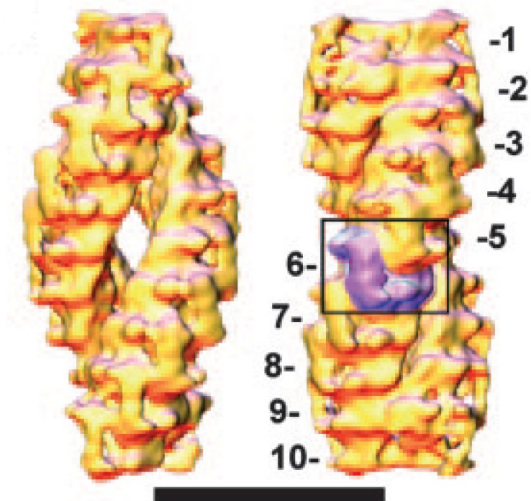
Automatic

Single-particle method



Tripeptidyl-peptidase II
(TPP II)

courtesy of B. Rockel, Martinsried



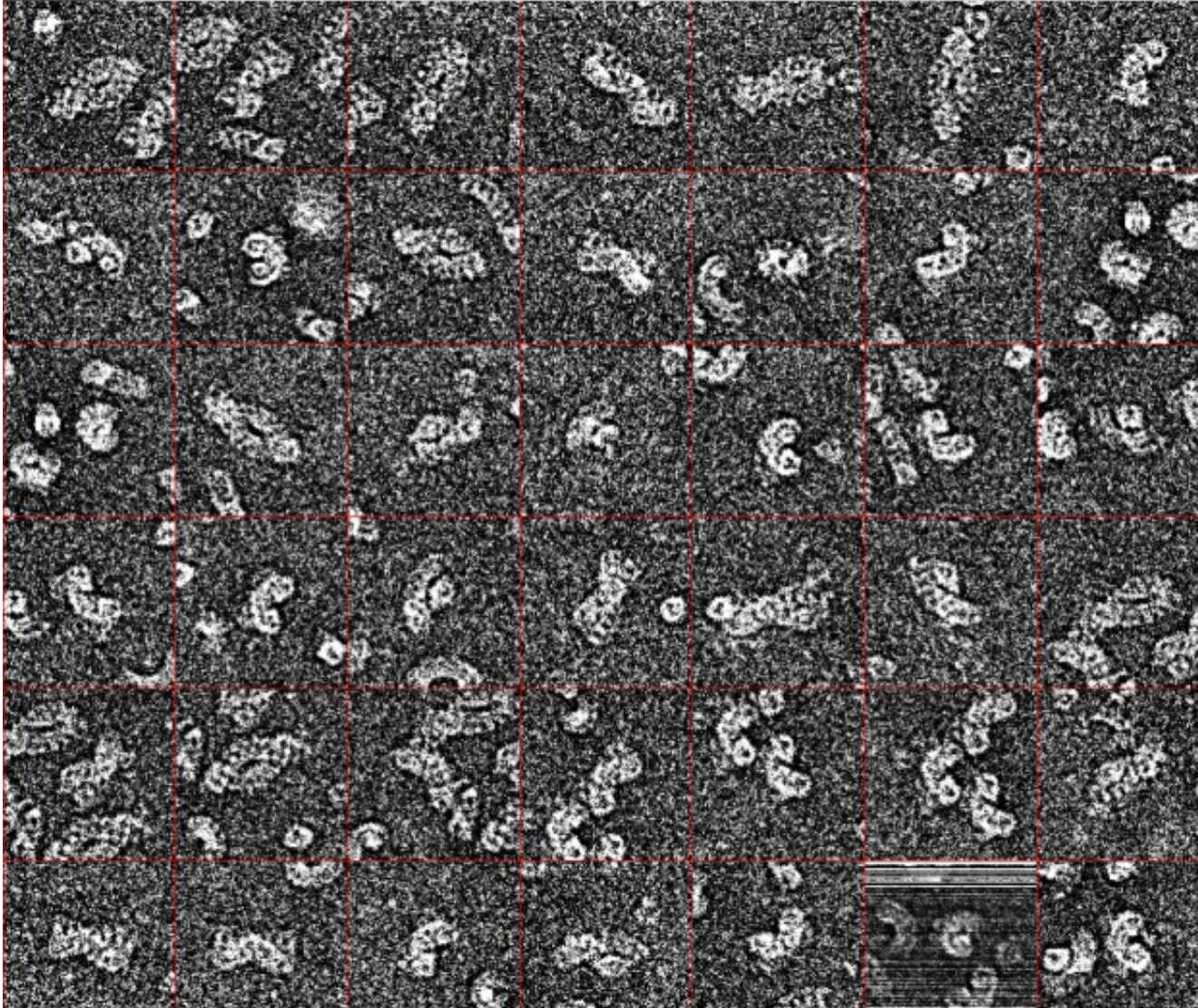
Single-particle analysis

- ▷ Obtain high resolution by computational averaging of images
- ▷ Assemble 2D projection views into a consistent 3D model
- ▷ Problem:

Orientations of views are unknown
→ hidden variables problem
EM algorithm

- ▷ Current solution:
ad hoc algorithm (EMAN)

Single particle stack

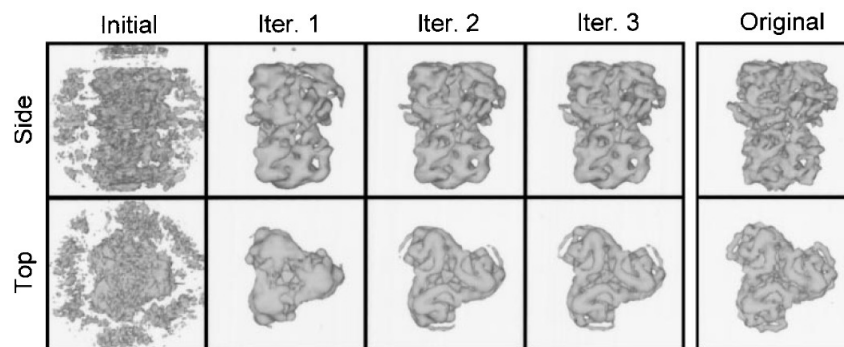
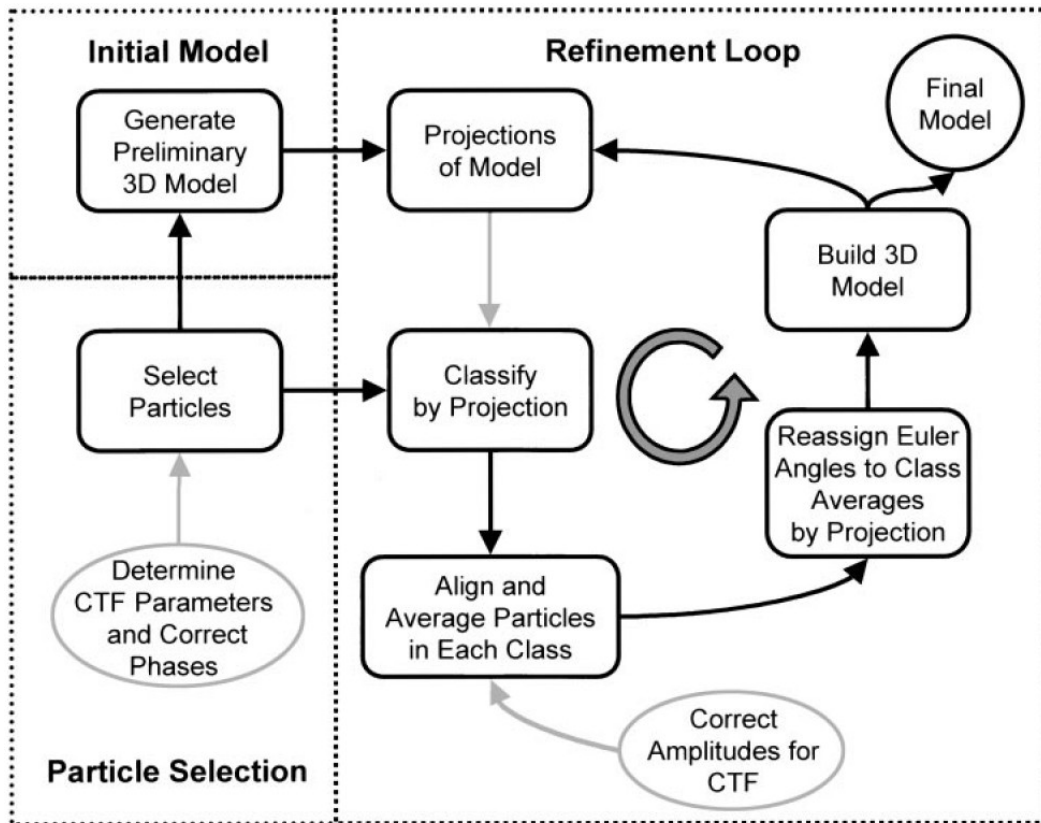


Typically several 1000 to 50000 individual images

Problems:

- Automatic selection of particles
- Alignment
- Classification
- Angular assignment
- 3D reconstruction

EMAN algorithm



EMAN single-particle reconstruction code

Steve Ludtke, Baylor College, Houston, TX

Ad hoc algorithm:

- No proof of convergence
- Local optima?

from: S. Ludtke, *J. Struct. Biol.*, 1999

EM algorithm

Parameter estimation with hidden variables:

Joint probability distribution:

normal distribution

$$p(I, \phi | V) = \mathcal{N}(I - P_\phi V)$$

One parameter (to be estimated):

$V = 3D$ volumetric density

Two random variables:

$I =$ Set of projection images (**observed**)

$\phi =$ Set of projection angles (**hidden**)

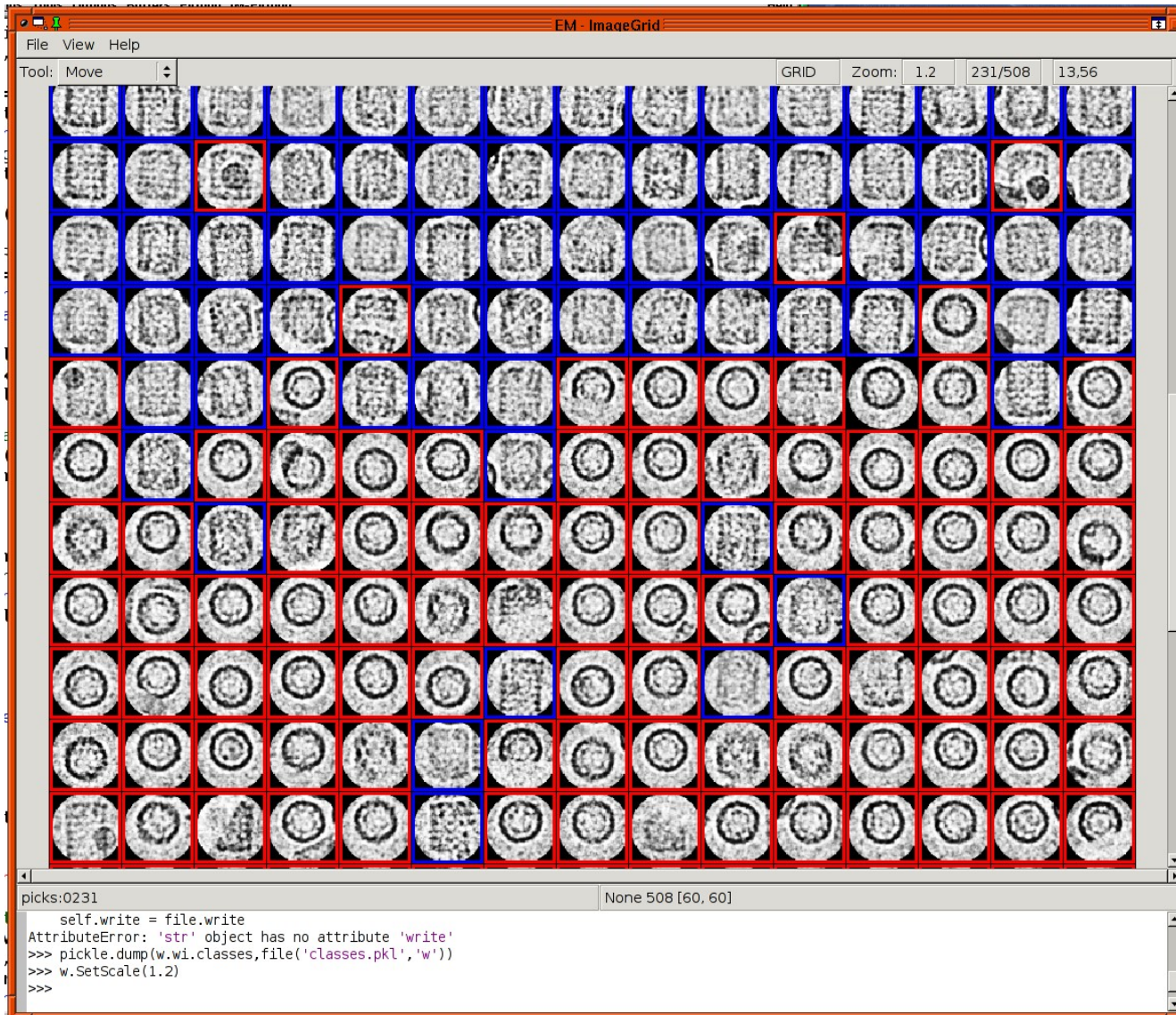
Particle picking

- ▷ Problem:
 - ▷ Views are randomly distributed on images
 - ▷ Must pick regions with particles from image
- ▷ Difficulty: high noise → simple template matching does not work
- ▷ Approach:

Initial picks by linear correlation

Use a Support Vector Machine (SVM) to select for correct particles according to a manually chosen data set

Picking by template matching



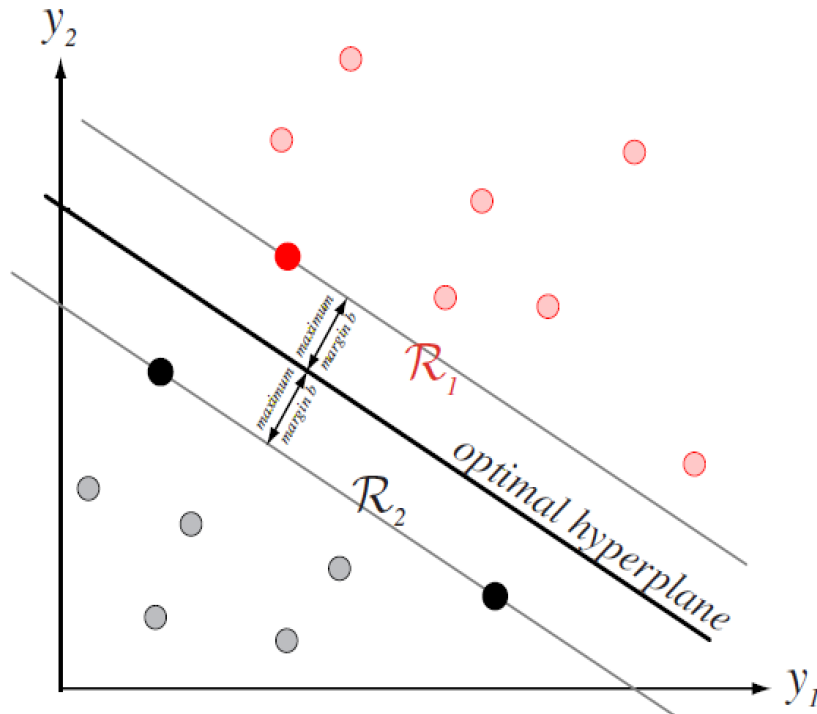
Picking by
linear correlation

many
mis-picks

Apply **SVM**
to pixel vector
(reduced) of
the images

*Coloring: training
data set*

Support Vector Machines



From Duda et al., *Pattern Classification*

Machine Learning:

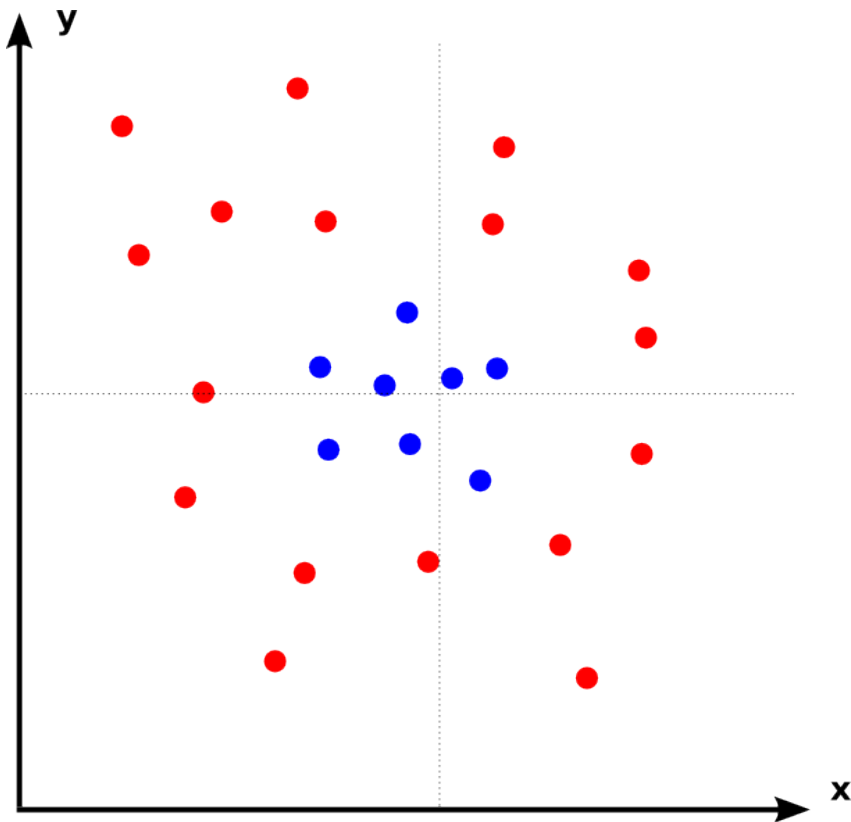
- Training (**vs rules**)

Support Vector Machine:

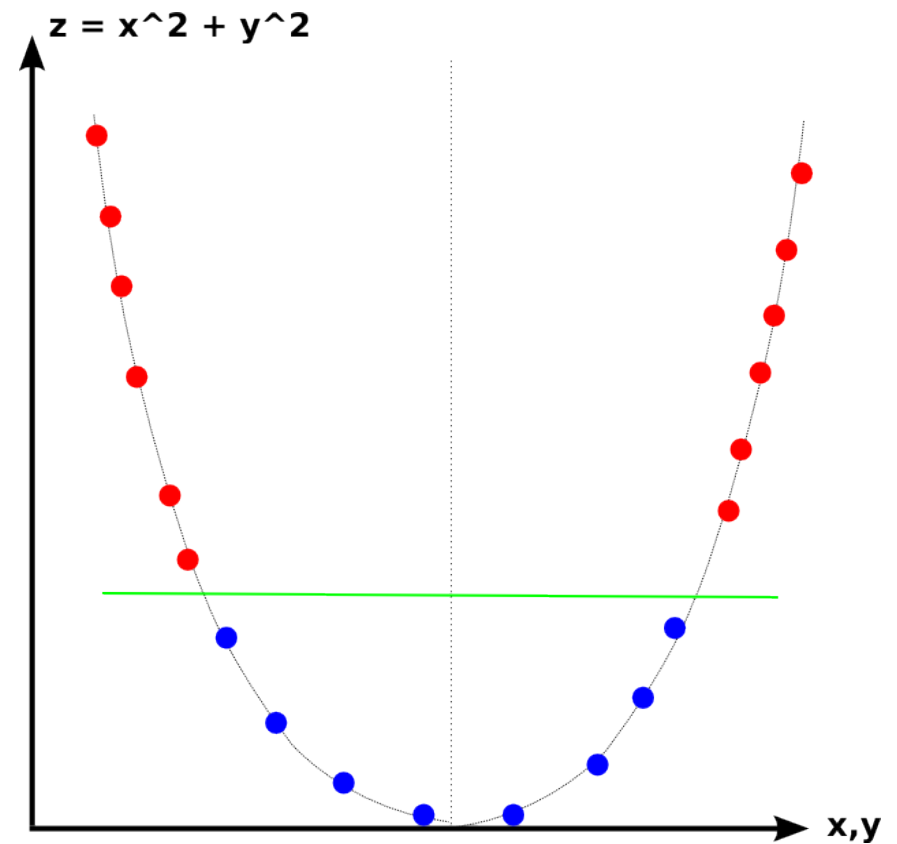
- Linear classifier
- Extended to higher polynomials
- Efficient calculation of the separating hyperplane by duality transform

Non-linearity

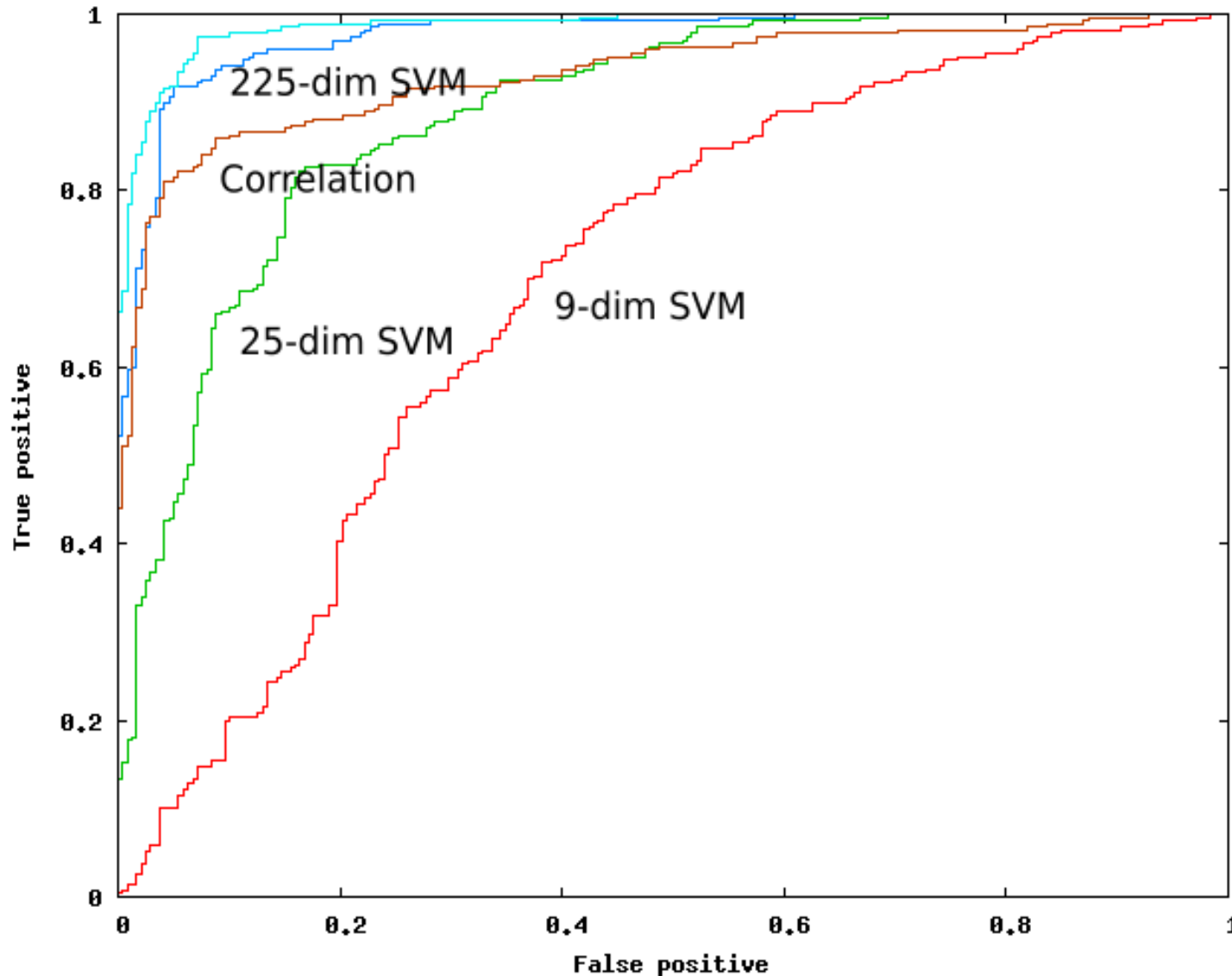
Linearly inseparable



Linearly separable
after introduction of pseudo-variable



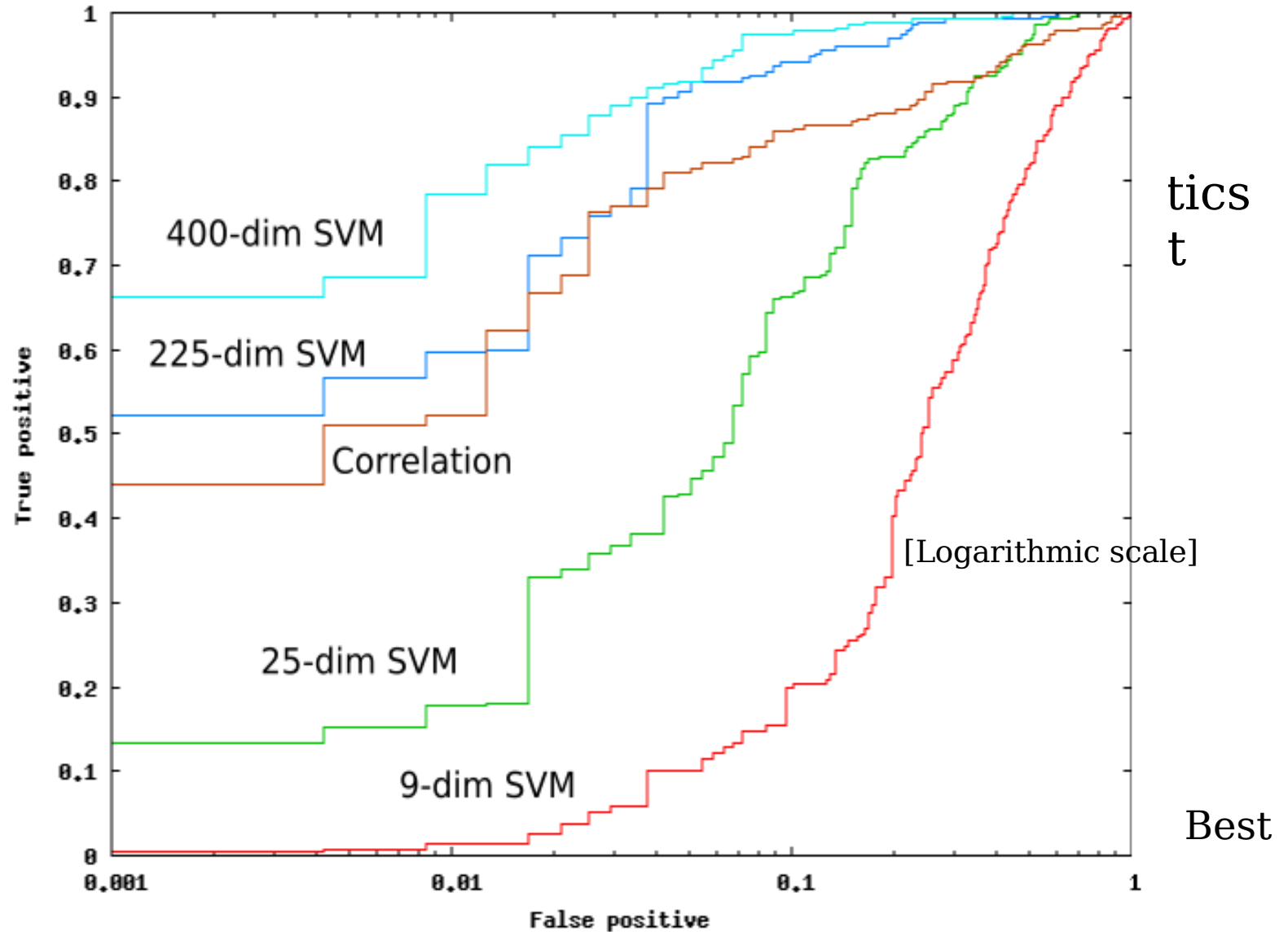
Improving picking using SVMs



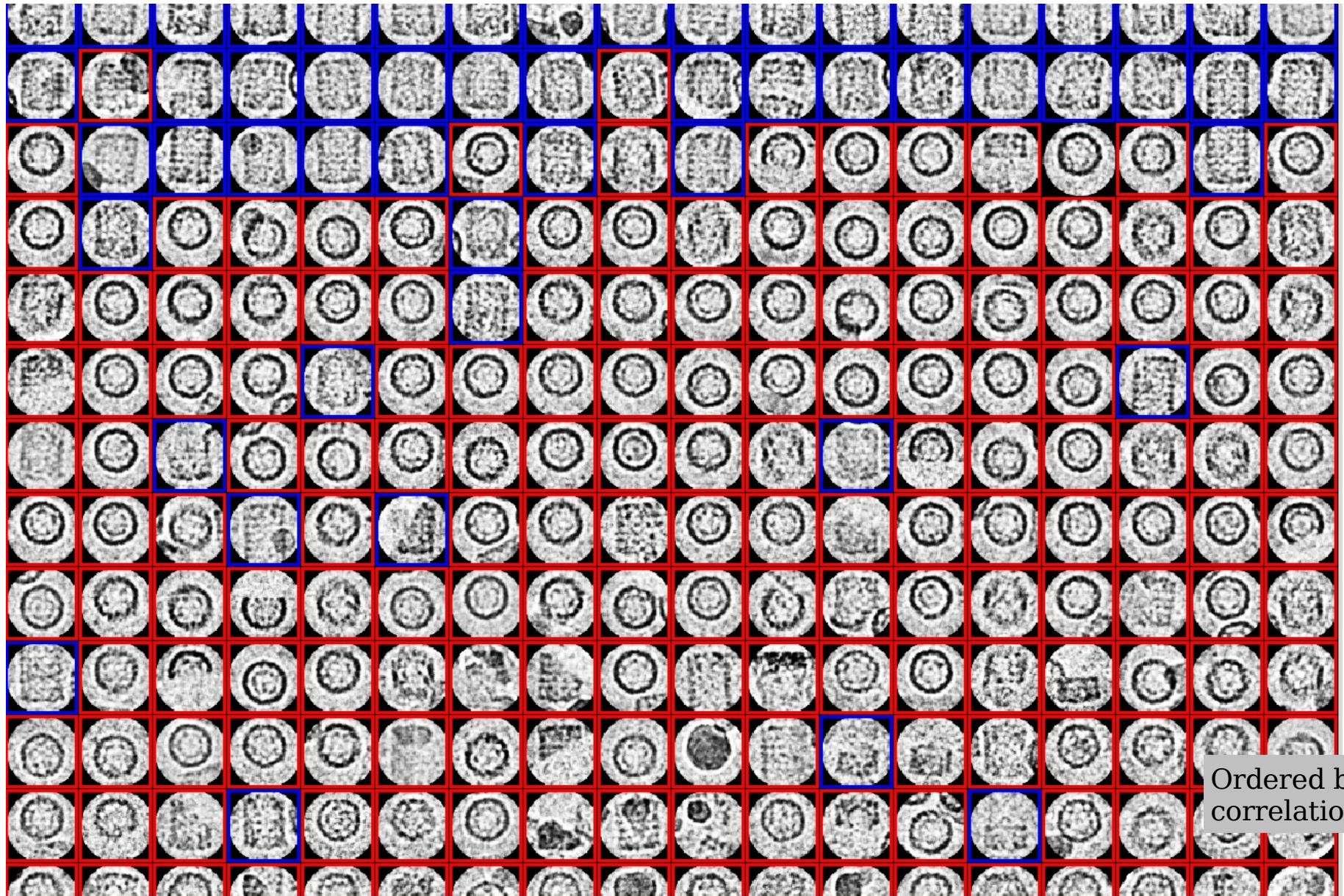
Receiver operating characteristics for different feature set sizes

M. Tacke, C. Best
2006

Improving picking using SVMs



Picking result



Ordered by
correlation value

Model-free particle classification

- ▷ How can we sort the views of a particle according to the viewing angle (elevation, azimuth) ?
- ▷ Answer: Similar angles → similar images
- ▷ Probabilistic model:

$$P(M|\phi; M_0, \phi_0) = \left(\frac{1}{2\pi\kappa(|\phi - \phi_0|)} \right)^{D/2} \exp \left(-\frac{|M - M_0|^2}{2\kappa(|\phi - \phi_0|)^2} \right)$$

Angular distance-to-similarity kernel

- ▷ Probability for an image M given an assigned angle ϕ , a reference image $M^{(0)}$, and a reference angle $\phi^{(0)}$:

Gaussian with a width that gets wider when the images are farther apart.

Self-organizing point map

- ▷ Joint probability distribution:

$$P(\{M^{(n)}\}|\{\phi^{(n)}\}) = \prod_{i=1}^N P(M^{(i)}|\{M^{(i')}, \phi^{(i')}\})$$

- ▷ Maximum-likelihood principle → Hamiltonian:

$$-\ln L(\phi) =$$

$$\sum_{n,m} \left(\underbrace{\frac{D}{2} \ln 2\pi\kappa(|\phi^{(n)} - \phi^{(m)}|)}_{\text{Attractive force}} + \underbrace{\frac{|M^{(n)} - M^{(m)}|^2}{2\kappa(|\phi^{(n)} - \phi^{(m)}|)^2}}_{\text{Repulsive force}} \right)$$

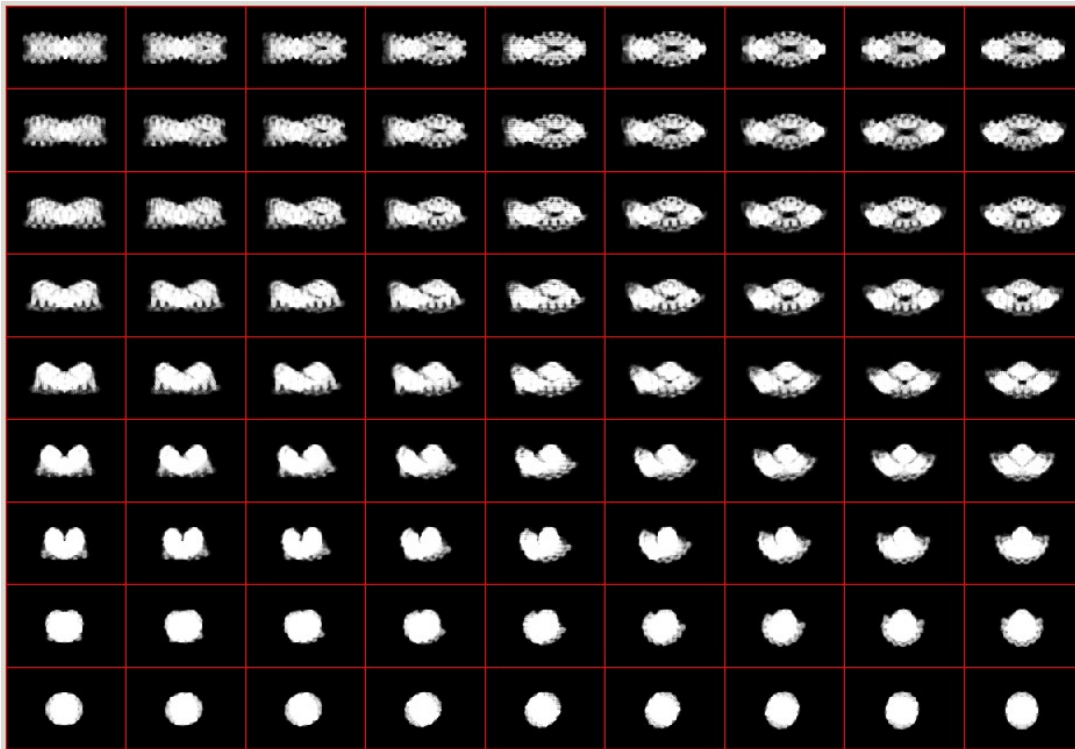
Attractive force

Repulsive force

Point-to-point potential → multidimensional scaling

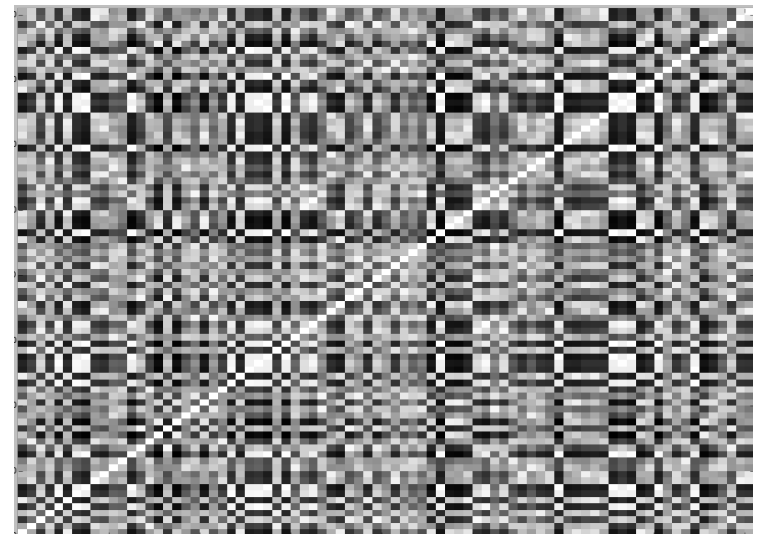
- ▷ Gradient descent solution

Similarity matrix



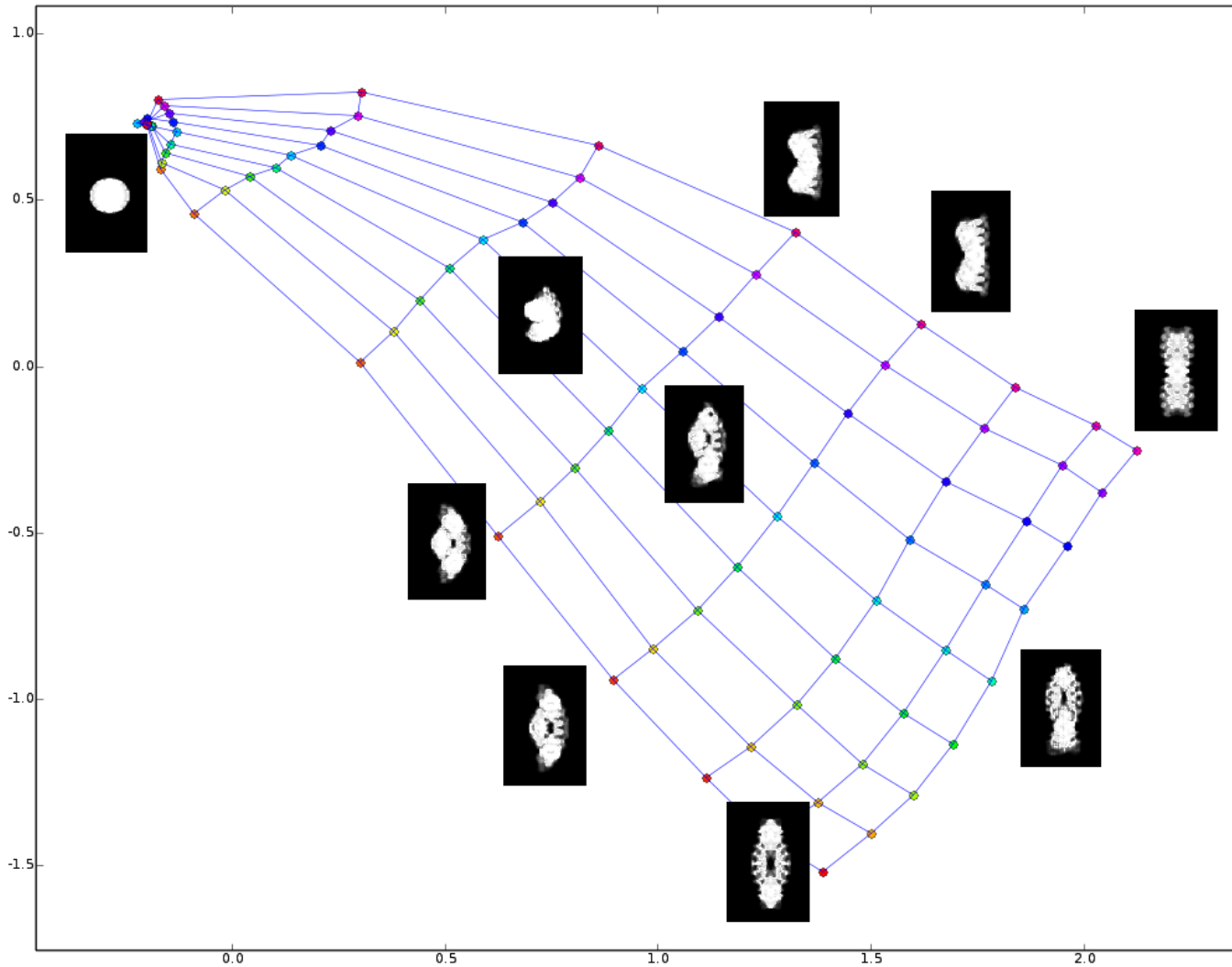
9x9 projections
of TPP2

Correlation matrix:



Pairwise correlation
max. over translations and
rotations

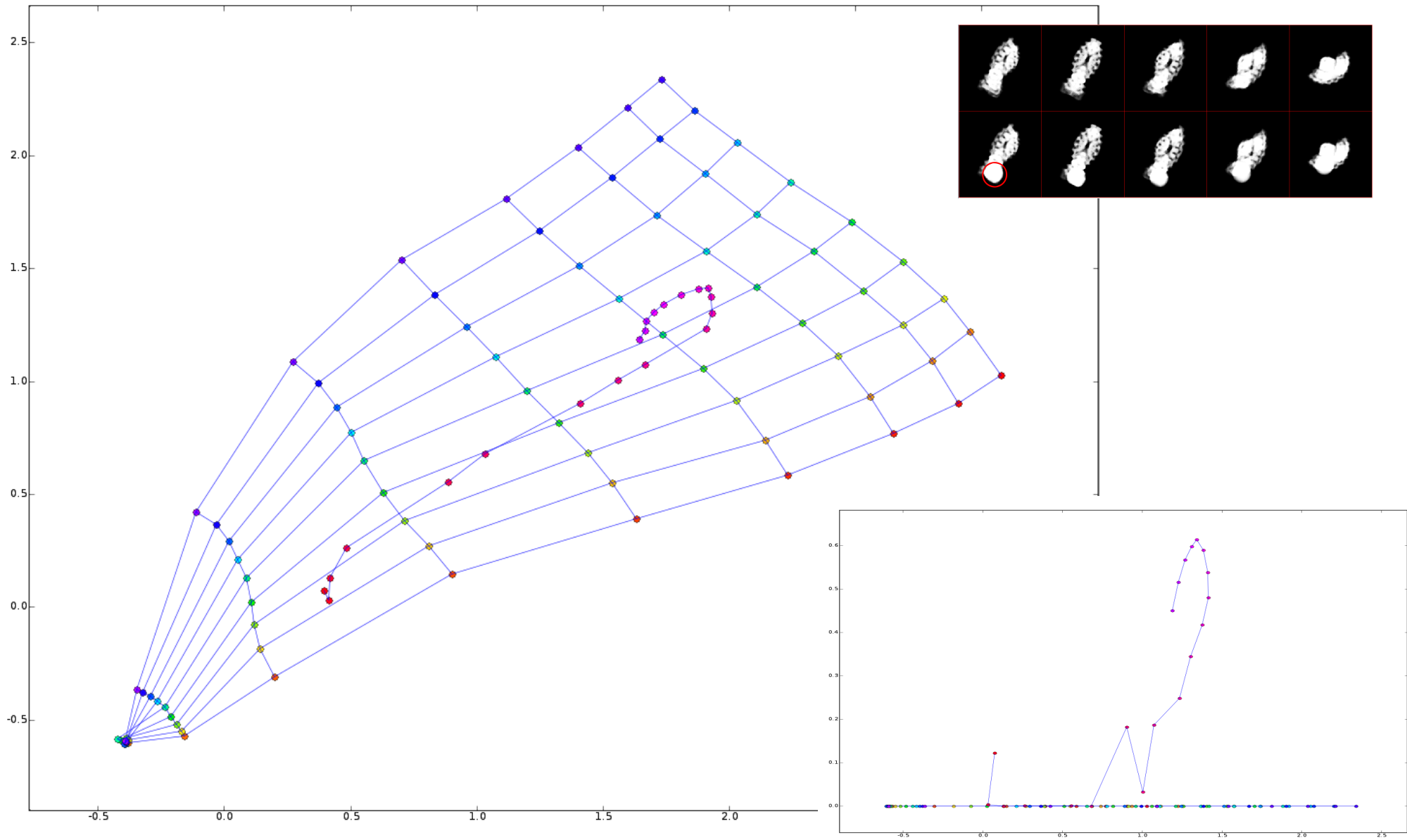
Result



Good
representation
of original
distribution of
viewing angles

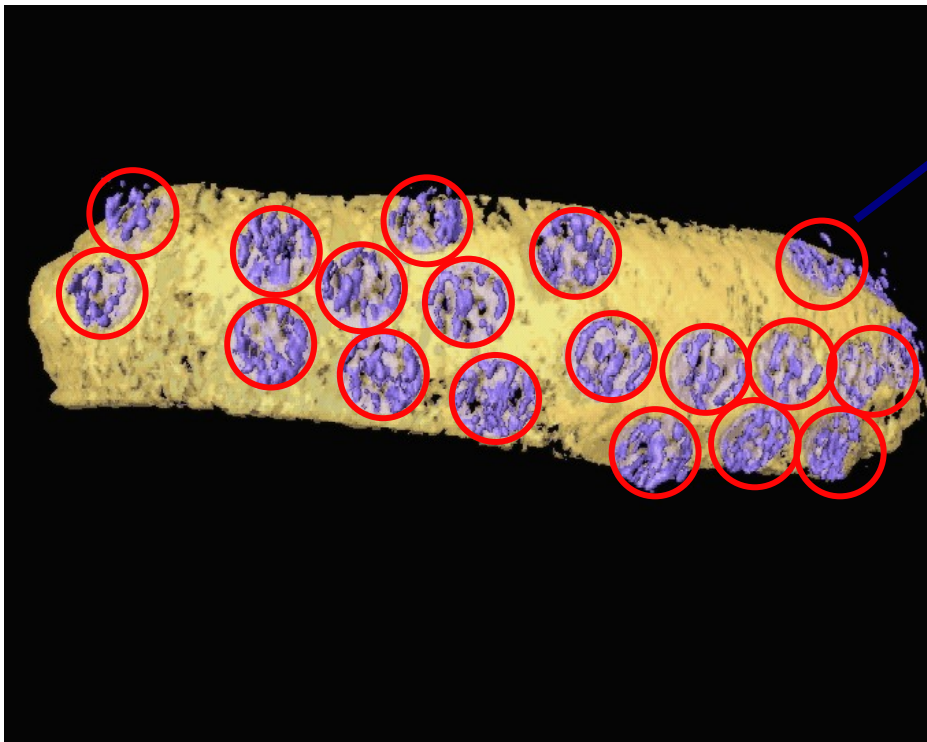
Good as an
initial model
for iterative
refinement

Tomographic classification



3D averaging

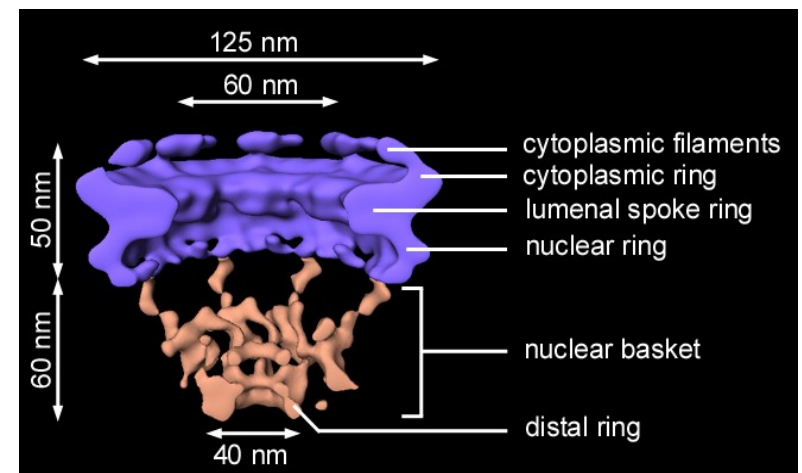
Nuclear pore complex



Matching

Aligning

Averaging

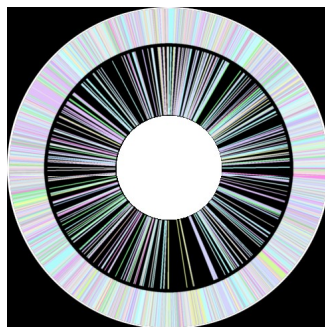


Beck *et al.*, *Science*, 2004

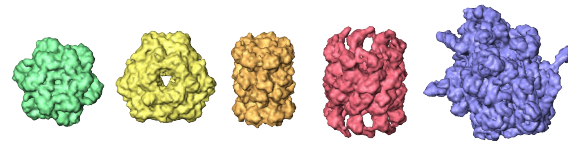
3D reconstruction

“Visual proteomics”

- ▷ Identify proteins in cryo-electron tomograms of intact cells or cryo-sections
- ▷ Pattern matching against a template library
- ▷ Map protein interaction landscape in cells
- ▷ High-throughput pipeline

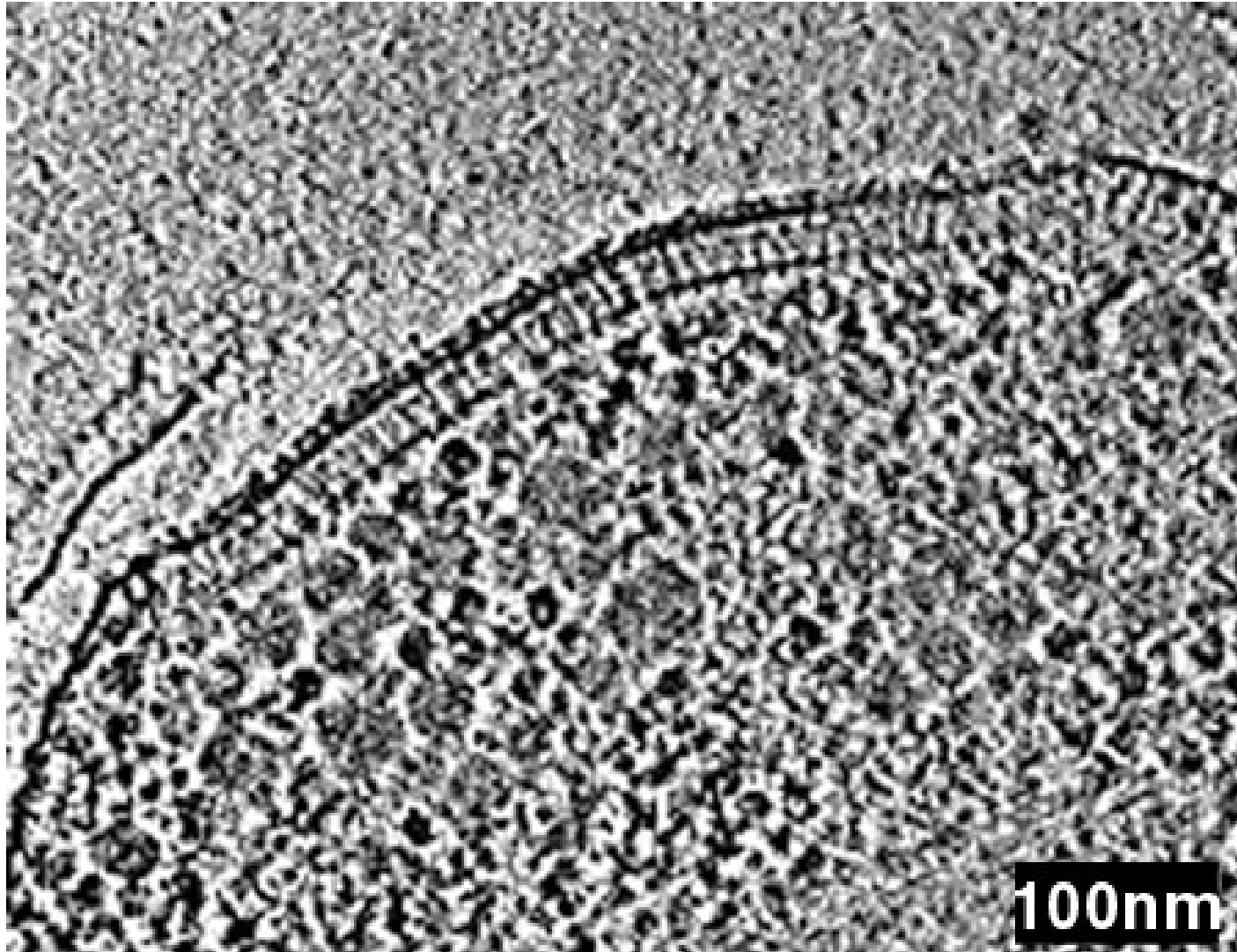


Genome / proteome



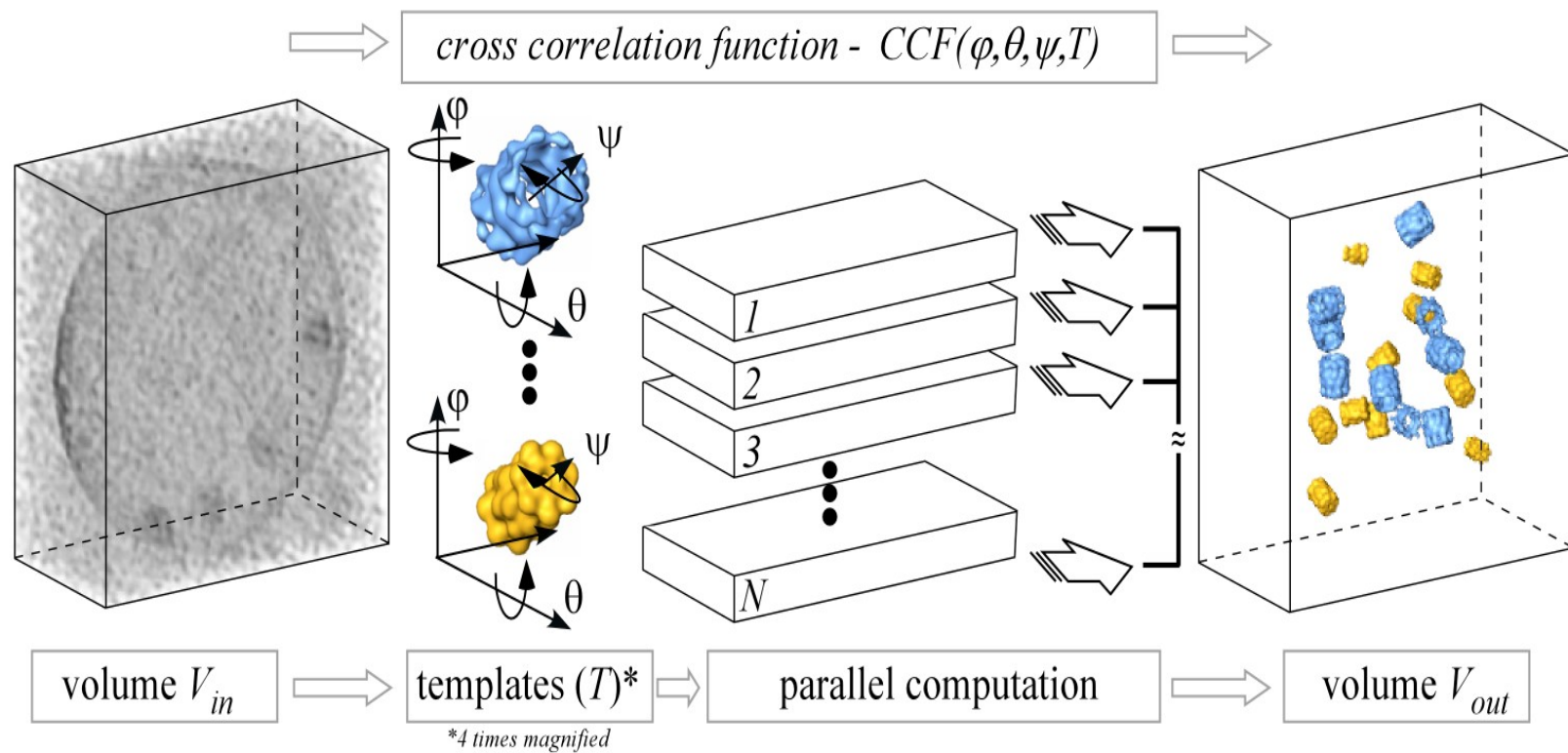
Template library

Macromolecular crowding

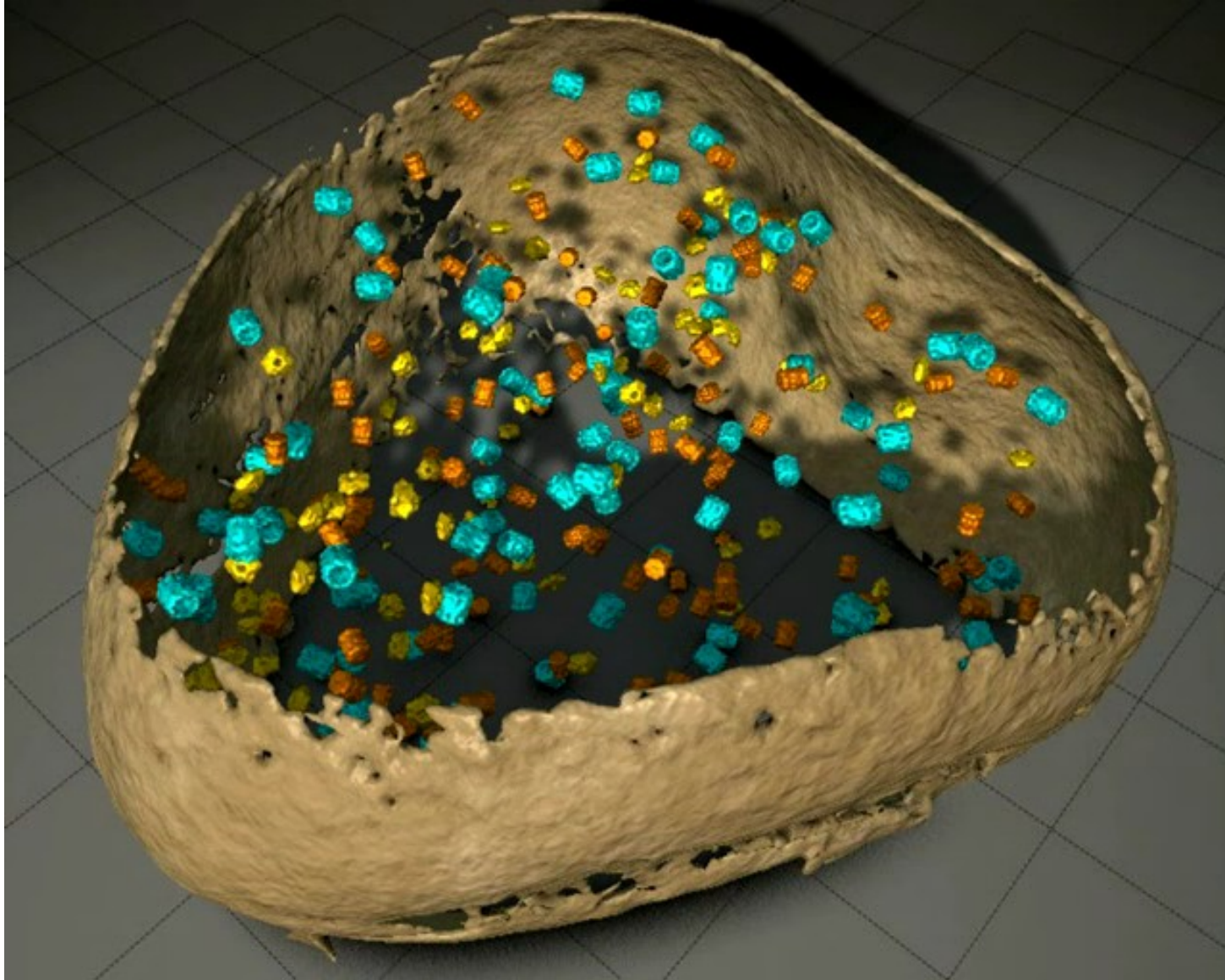


Visual proteomics

Computational pipeline:



“Visual proteomics”

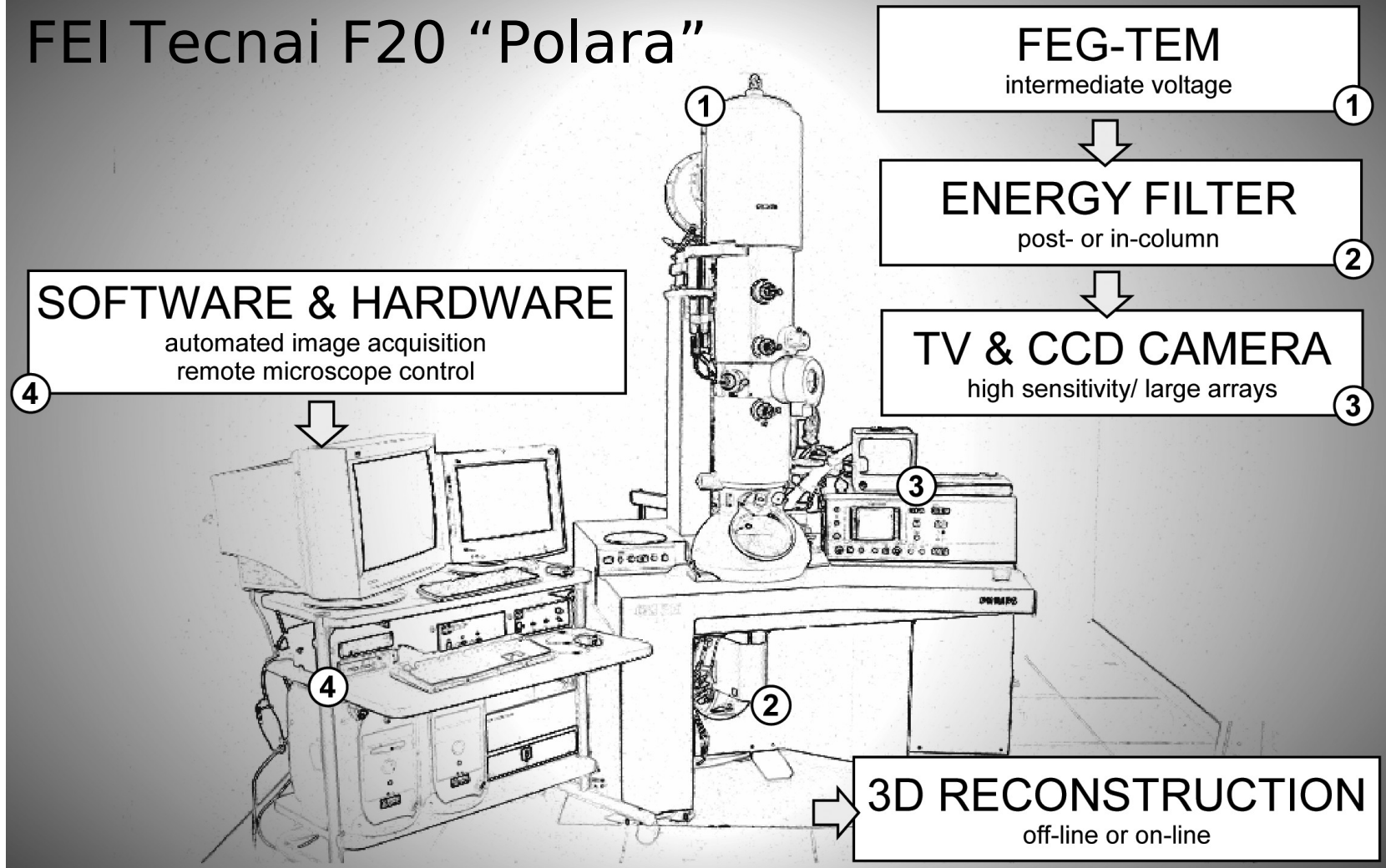


Conclusions

- ▷ Cryo-electron tomography allows **molecular resolution** imaging of cells in **near live** conditions
- ▷ Depends heavily on computational methods for **reconstruction, denoising, and segmentation.**
- ▷ Single-particle averaging methods (both 2D and 3D) requires sophisticated **pattern recognition, classification, and clustering methods**
- ▷ Future influences:
 - ▷ Probabilistic modelling
 - ▷ Monte Carlo algorithms
 - ▷ Machine learning

Electron microscope

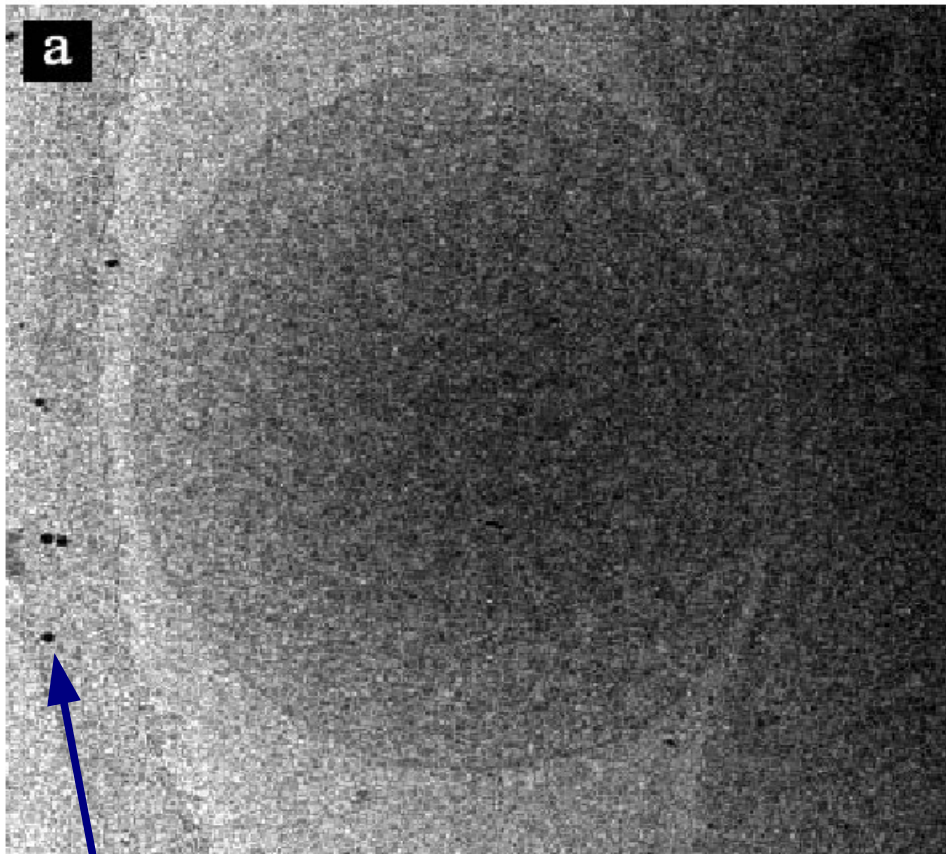
FEI Tecnai F20 "Polaris"



Price tag: around \$1.5M

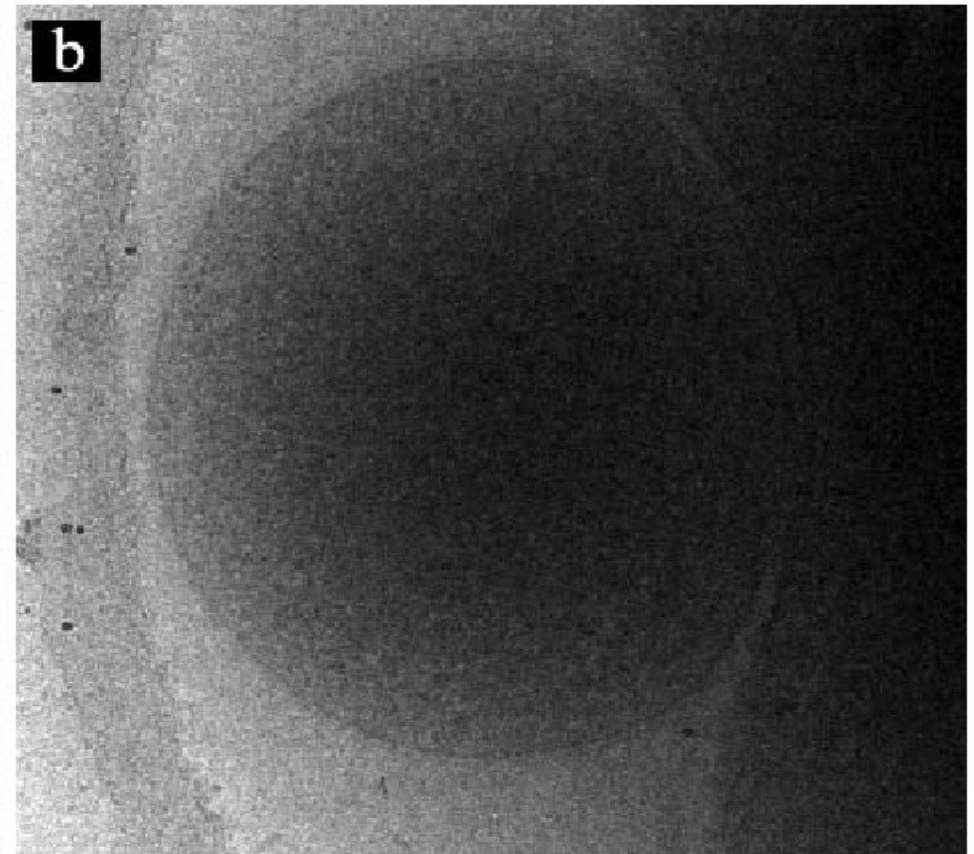
Electron micrographs

Thermoplasma acidophilum



No energy filtering

Gold markers for alignment



With energy filtering

Image: C. Kofler, Martinsried

Molecular scale imaging

- ▷ Light microscopy: limited by wavelength $O(100\text{nm})$
- ▷ Alternative probes:
 - ▷ X-rays
 - ▷ Short wavelength \rightarrow atomic resolution ($< 0.1\text{nm}$)
 - ▷ Low cross-section, difficult optics \rightarrow need crystals
 - ▷ Electrons:
 - ▷ Particle-wave duality, de Broglie wavelength
 - ▷ Charged massive particles \rightarrow high cross section
 - ▷ But high damage to specimen \rightarrow low dose
 - ▷ **High noise**

EM algorithm

Estimation step:

$$Q(V|V_0) = \sum_{\phi} p(\phi|I, V) \log p(I, \phi|V)$$

Minimization step:

$$V^{(n+1)} = \max_V Q(V|V^{(n)})$$

Differences to ad hoc algorithm:

- $p(\phi|I, V)$ is unity (for assigned angle) or zero (otherwise)